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Investigations into the Non-Mevalonate Isoprenoid Biosynthesis Pathway's First Two Enzymes utilizing Hybrid QM/MM Techniques

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Investigations into the Non-Mevalonate Isoprenoid Biosynthesis Pathway’s First Two Enzymes utilizing Hybrid QM/MM Techniques

by

Justin K. White

A dissertation submitted in partial fulfillment of the requirements for the degree of Doctor of Philosophy Department of Chemistry College of Arts and Sciences University of South Florida

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Keywords: QM/MM, computational, 1-deoxy-D-xylulose 5-phosphate synthase, 1-deoxy-D-xylulose 5-phosphate reductoisomerase

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Dedication

I dedicate this to my constant companion and greatest supporter, Molly Burges.
Acknowledgments

To my family, I would like to express my unending gratitude for the support shown me over the past many years. As I am slogged through this process and begun to confront personal issues, you have been along side me.

My friends of whom, I consider my extended family. You have thought better of me than I have. For that and your undying faith in my potential, I will be eternally grateful. You are always there when I need you, no matter how long it has been since we last spoke.

The love of my life already got the dedication but she deserves so much more. My new wife is always in my corner. Trying to look out for me, even when I don’t care about what happens to me. She is my love and rock. She has been there through it all.

I am grateful to my Ph.D. committee of Lee Woodcock, David Merkler, Wayne Guida, and Chair Yu Chen for their guidance and patience over the years. Though I might have pushed the patience portion too much.

My labmates through the years: Christi Whittington, Sai Vankayala, Fiona Kearns, Phillip Hudson, Yura Pevzner, and Jackie Hargis. There might be others I am forgetting but still know that each of you have helped me along the path, if by no other way then being there for me to rant at for a while.

Lastly, I am grateful for the opportunities and guidance provided by Lee and Dave over these many years. I hope I haven’t caused you too many causes for stress and know that I do truly appreciate every bit of guidance and patience along the way.
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Abstract

Molecular drug design begins with the identification of a problem to solve. This work identifies the growing resistance among human pathogens to current treatments. Once the problem is identified and understood, solutions must be proposed. This one is straightforward, we need new antimicrobial drugs. More specifically, we need to identify novel targets to inhibit. A large portion of antibiotics focus on disruption of macromolecular production while only a few target metabolic systems. Finally, you need to propose solutions based on the information gathered. In order to avoid existing resistance, it is important to avoid the macromolecular route and focus on metabolic enzymes. Preferably, the pathway would have little overlap or similarity with pathways found in the treatment organism. With this in mind, the non-mevalonate (NMA) pathway poses as a very good target for drug design. Many pathogens have been found to be strictly dependent on this pathway while it is absent in humans. Additionally, fosmidomycin has already been shown to inhibit this pathway. Initially thought to just inhibit the 1-deoxy-D-xylulose 5-phosphate (DXP) reductoisomerase (DXR), it has been shown to inhibit several enzymes along the path to a lesser extent. Ideally, this could be repeated or improve upon for future drug design.

With this in mind, the initial stages of the first two enzymes of the NMA pathway were examined utilizing quantum mechanical/molecular mechanical (QM/MM) techniques. The first enzyme was DXP synthase (DXS), which catalyzes a transketolase-like condensation of pyruvate and glyceraldehyde-3-phosphate to produce DXP. DXS and other transketolases are dependent on the thiamine diphosphate (TDP) cofactor, which must be deprotonated of the imidazolium C2 atom producing a highly reactive ylide. A tautomerization occurs prior to this deprotonation to prime the pyrimidinium ring N4 atom to perform the C2 abstraction. The question at hand was the identity of a general base
to perform the N4 abstraction. The results favored a water-mediate mechanism with a higher than usual $\Delta E^\ddagger$ of 22.7 kcal/mol. An observation pertaining the tautomerization pertained to the aromaticity of the pyrimidine ring. Upon further investigation, aromaticity was found to play a significant role in the $\Delta E^\ddagger$ observed. Aromaticity might contribute 14.2 kcal/mol to the barrier height. This high energy would drive the reaction forward producing the ylide.

Investigation of the DXR enzyme followed this work. Initially, the work was going to focus on the 2 mechanisms proposed for activity, $\alpha$-ketol rearrangement and retro-aldol/aldol mechanism. Subsequent publications involving secondary kinetic isotope effects (KIEs) add to the pile of evidence supporting the retro-aldol/aldol mechanism. So the project was retooled to investigate the energetic differences between two metal binding modes. The results of this work support a metal coordination across the C3-C4 bond, which eventually extends coordination to include the C2 oxygen. This conformation was help explain the tight binding effecting observation of the putative intermediates (transition states) and aldehyde intermediate. Additionally, as the C2-C3 mode consistently transfers a proton to the phosphate group of DXP or produces an elongated C-O bond, the C2-C3 mode would not be favorable.

Further investigations of these enzymes (e.g. completing the step begin, continuing through the reaction) could provide further illumination into the mechanism of action and possibly reveal new avenues of drug design. Examining the enzymes downstream in the NMA pathway might provide details of interest. Of particular interest is the radical reaction proposed for HDR/IspH. The final step of the pathway produces IDP and DMADP in a 4:1 proportion, which corresponds to the general system requirements for production of the long chain, branched isoprenoids. It would be interesting to compute the mechanism to see if energetics could provide further insights. Additionally, normal mode analysis coupled with vibrational subsystem analysis could identify allosteric sites for feedback sensitivity.
Chapter 1

Introduction

1.1 Isoprenoids

Accounting for nearly 60% of natural product diversity, isoprenoids (or terpenoids), with 55,000 known compounds, comprise the largest family of natural products\textsuperscript{56,59}. Many of these compounds serve important biological functions. The lipid-soluble vitamins (A, D, E, And K) and cholesterol are some of the most common examples\textsuperscript{161}. Cholesterol is subsequently utilized as a biosynthetic precursor of various steroid hormones, including glucocorticoids, estrogens and androgens. Some synthetic analogs are used in many therapeutic applications. All isoprenoids are derived from two phosphate C\textsubscript{5} isoprene building blocks, isopentenylallyl diphosphate (IDP) and dimethylallyl diphosphate (DMADP). The diversity of isoprenoids comes from number of IDP and DMADP molecules strung together. The string of isoprenes in combination with functional groups such as ketones, aldehydes, alcohols, peroxides, ethers and esters contribute to the considerable diversity amongst the family of natural products\textsuperscript{47}.

The family is roughly divided into six major categories (Figure 1.1. These categories are based on the number of isoprene units linked. Monoterpenoids contain two isoprene units and therefore have a ten carbon skeleton. These are the major component of the fragrant oils from leaves, flowers and fruits (e.g, limonene and nerol). Sesquiterpenoids consist of three isoprene units to form 15 carbon cyclic and acyclic compounds. The next category is called diterpenoid and are composed of 20 carbon atoms derived from geranyl
<table>
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<th>Compound</th>
<th>Description</th>
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<tr>
<td>Limonene</td>
<td>Cyclic terpene utilized for citrus flavor by the cosmetic industry</td>
</tr>
<tr>
<td>Abscisic Acid</td>
<td>Plant hormone required for development processes</td>
</tr>
<tr>
<td>Retinol</td>
<td>Required for vision and derived from vitamin A</td>
</tr>
<tr>
<td>Ubiquinone</td>
<td>Utilized as part of the electron transport chain for cellular respiration</td>
</tr>
<tr>
<td>Cholesterol</td>
<td>Precursor to bile acids, steroid hormones and vitamin D; required for cell membrane fluidity</td>
</tr>
<tr>
<td>Vitamin E</td>
<td>Antioxidant to reduce the production of reactive oxygen species formation during lipid oxidation</td>
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geranoil diphosphate (a 10 carbon unit). Vitamin A, phytohormone and tetrahydrocannabinol are a few of the more well characterized examples of diterpenes. Ophiobolin A, a fungal metabolite, is an example of the next category, sesterterpenoids. These are derived from 25 carbon framework. Cholesterol are a member of the triterpenoids which are desired from the squalene precursor. Carotenoids are comprised of eight isoprene units to make forty carbon chains with conjugated double bonds. Carotenoids utilize the
absorption properties arising from their conjugated structures to assist in photosynthesis and the prevention of photo-oxidative cellular damage.

1.2 Pathways for Isoprenoid Biosynthesis

1.2.1 Mevalonic Acid Dependent Synthesis of Isoprenoid Building Blocks

The first pathway for isoprenoid biosynthesis was discovered based on interest in cholesterol for the obvious health related interests. During the investigation of cholesterol, researchers discovered deuterium integration originating from labeled acetate via the IDP unit which meant IDP was the direct precursor to cholesterol\textsuperscript{16}. Subsequent studies lead to the discovery and characterization of the mevalonate (MVA) pathway in the 1950s named after the key intermediate (3R)-3,5-dihydroxy-3-methylpentanoic acid (mevalonic acid, MVA). For the following decades, the MVA pathway dominated this area of research as it was thought to be the sole route for IDP and DMADP synthesis in living systems. The work in this area lead to a nobel prize in physiology for Lynen and Bloch in 1964, and in Chemistry for Cornforth in 1975\textsuperscript{15,40}.

As figure 1.1 illustrates\textsuperscript{130}, the initial step of the MVA pathway is the production of acetoacetyl-CoA via the condensation of two acetyl-CoA molecules catalyzed by acetyl-CoA acetyltransferase. A third acetyl-CoA molecule is attached to the acetoacetyl-CoA to produce 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA) catalyzed via an aldol reaction performed by HMG-CoA synthase. HMG-CoA is reduced by two equivalents of NADPH performed by HMG-CoA reductase producing MVA. The HMG-CoA reduction is the rate-limiting step of this pathway, thus MVA production is the rate-limiting, or key, intermediary step in the pathway. Two consecutive phosphorylations performed by mevalonate kinase and phosphomevalonate kinase produces mevalonate-5-diphosphate (MDP). An ATP-coupled decarboxylation catalyzed by MDP decarboxylase yields IDP\textsuperscript{130}. The IDP isomer, DMADP, is produced via two structurally unrelated IDP:DMADP isomerases
Figure 1.1: An illustration of the complete Mevalonate and Non-Mevalonate pathways culminating in the production of IDP and DMADP.
1.2.2 Mevalonate-Independent Synthesis of IDP and DMADP

Decades following the MVA pathway discovery, further isotopic labeling studies were performed to trace the fate acetate in producing lycopene, hopanoids, taxol and sterols. The distribution of the labeled material in the subsequent terpenoids was inconsistent with a single source of isoprenic production. Following from these results, several independent research groups discovered a mevalonate-independent (or non-mevalonate, NMA) pathway in eubacteria, green algae, and higher plants. Rohmer et al. identified the conversion of pyruvate to 1-deoxy-D-xylulose 5-phosphate (DXP) as the first step of the NMA pathway by following the incorporation of $^{13}$C-labeled pyruvate or glycerol into ubiquinone. Additional independent studies carried out by Arigoni et al. traced the incorporation of [1-$^{13}$C]- and [2,3,4,5-$^{13}$C$_4$]-DXP into the formation of β-carotene, lutein, phytol and sitosterol in cell cultures of Catharanthus roseus and demonstrated the involvement of DXP in the NMA pathway. The studies conducted by Arigoni et al. provided further insight into the compartmentalization of isoprenoid synthesis, as well as, the description of the rearrangement proposed by Eisenreich et al. In order to elucidate the NMA pathway (Figure 1.1, further labelling studies were performed and revealed the pathway to be composed of 7 enzymes that catalyze 8 reactions. Background on each of the enzymes found in the NMA pathway can be found in the following paragraphs.

1-Deoxy-D-xylulose 5-Phosphate Synthase (DXS)

The aforementioned conversion of pyruvate to DXP is performed by DXP synthase (DXS) and employs glyceraldehyde-3-phosphate (G3P) (Figure 1.2). DXS is a member a large family of enzymes dependent upon the cofactor thiamine diphosphate (TDP$^{176}$). Particularly, the structure and reaction catalyzed are highly reminiscent of members of the
subfamily of transketolases (TKs). Structurally, DXS is similar in composition to other members of the subfamily. Each monomer is composed of 3 subunits (I, II and III). In solution, DXS is more commonly found in a homodimer that is functionally significant as each active site communicates with the other. The formation of the active site pocket distinguishes DXS from other members of the TK subfamily. The active site in other TKs exists between domain I of one monomer and domain II of the other monomer II in a twisted conformation that arises via formation of the homodimer. In contrast, the DXS active site resides in a pocket between domains I and II of the same. Despite this distinction, several key residues remained highly conserved with the rest of the subfamily.

Mechanistically speaking, DXS corresponds to the α-hydroxyketone (acyloin) condensation and proceeds via a mechanism highly analogous to other TKs. Prior to substrate binding, the TDP cofactor undergoes a deprotonation of the thiazolium ring forming a carbanion ylide necessary for enzymatic. The usually high pKₐ of the thiazolium proton makes this reaction highly unlikely in solution. The active site of DXS and other TKs binds the TDP into a energetically strained ‘V’-shaped conformation; which brings the N4 of the pyrimidine ring into proximity of the C2 thiazolium proton. The proximity of nitrogen and strained structure lowers the pKₐ significantly. Therefore, the carbanion ylide is free to perform a nucleophilic attack on the pyruvate. Subsequently, the electrophilic iminium acts as an electron sink during decarboxylation and results in a carbanion/enamine. The enamine performs a second nucleophilic attack on carbonyl carbon of glyceraldehyde-3-phosphate (G3P). A final deprotonation step releases the new

![Figure 1.2: The abridged representation of the DXS catalyzed condensation of pyruvate with glyceraldehyde-3-phosphate to produce 1-deoxy-D-xylulose 5-phosphate.](image-url)
DXP molecule and regenerates the TDP ylide for further catalysis. The substrate binding mechanism of other TKs has been thought to work through either a ping-pong or sequential mechanism. Recent studies of DXS have suggested a random sequential mechanism thus illustrating a further distinction between DXS and other TKs. DXS does show a preferred order involving the formation of the C2α-lactylthiamin diphosphate (LTDP) intermediate as an unusually stable ternary complex of TDP and pyruvate. The hydrox-yaldehyde moiety of G3P was found to trigger and accelerate the decarboxylation which produces the enamine utilized in the second nucleophilic reaction.

1-Deoxy-D-xylulose 5-Phosphate Reductoisomerase (DXR)

The next step in the NMA pathway is actually the first committed step of the pathway. DXP reductoisomerase (DXR) catalyzes the conversion DXP into 2-C-methyl-D-erythritol 4-phosphate (MEP) with preferential dependence on NADPH as reducing agent and Mn$^{2+}$ as a divalent ion. The carbon-skeleton rearrangement in this reaction is thought to proceed via the aldehyde intermediate, 2-C-methyl-D-erythrose 4-phosphate (MEsP), which is subsequently reduced by NADPH (Figure 1.3). The idea behind the aldehyde intermediate arose due to similarities between DXR and ketol-acid reductoisomerase (KARI; EC 1.1.1.86); which catalyzes a similar rearrangement-reduction sequence in the conversion of 2-acetolactate to 2,3-dihydroxy-3-methylbutyrate. In both situations, the intermediate has never been directly observed as it is either tightly bound prior to reduction or in such low concentration due to it’s transient nature. An experiment by Rohmer and co-workers provided the strongest evidence in support of the MEsP intermediate. The researchers synthesized MEsP and demonstrated its kinetic competency with DXR in the presence of NADPH and Mn$^{2+}$ or Mg$^{2+}$. They also observed a 7% conversion of MEsP to DXP in the presence of NADP$^+$. 
Similar to DXS, DXR is most commonly found in a homodimer of the V-shaped monomers producing a saddle-like quaternary structure. The monomers can be further subdivided into three distinct domains. A dinucleotide-binding domain acts as a binding site for the NADPH cofactor is found in the N-terminal region of each monomer. The central domain harbors the catalytic portion of the enzyme and is responsible for the crucial conformational changes required for substrate binding and turnover. The central domain contains a highly flexible loop, which acts as a lid upon substrate binding creating a protected active-site cavity. The final domain is the C-terminal domain consisting of a four-helix bundle and is characterized by its flexibility. The role of this flexible domain is to aid in dimerization.\textsuperscript{118,151,199,200}

Despite the similarities between DXR and KARI, differences in amino acid sequences and crystal structures suggest different mechanisms for DXR.\textsuperscript{37,49,95} Three mechanisms were originally considered for DXR’s rearrangement of DXP to MEsP: 1) an α-ketol rearrangement, 2) a retro-aldolization/aldolization and 3) a sequential 1,2-hydride and 1,2-methyl shift.\textsuperscript{64} This last proposal was readily eliminated when the $^{13}$C-glucose incorporation studies failed to yield the appropriate labeled products. More over, 2-$^{13}$C and 3,4,5-$^{13}$C$_3$ labeled DXP experiments strictly yielded [2-$^{13}$C]- and [1,3,4-$^{13}$C$_3$]-MEP, respectively, which supports the rejection of the sequential shift mechanism.\textsuperscript{3,73,160} The α-ketol (sigmatropic) rearrangement occurs via the migration of the C3-C4 bond to form a C2-C4 bond in order to form MEsP, the aldehyde intermediate. This migration re-
quires a partial positive charge on the C2 atom which can be achieved via protonation or the ketol coordinating with the divalent metal ion. There is evidence to support the latter approach. The metal has been shown to be chelated by the hydroxy groups of DXP. A deprotonation of the C3 hydroxyl group of DXP is required for aldehyde formation. Thus, the deprotonation and bond-cleavage/formation would result in the MEsP. Alternatively, the retro-aldol/aldol reaction begins with the deprotonation of the C4 hydroxyl group followed by cleavage of the C3-C4 bond in a retro-aldolization. The result of this reaction is the formation of a hydroxyacetone enolate and glycoaldehyde phosphate. Subsequently, an aldol condensation will result in the same MEsP intermediate. Kinetic isotope effects (KIEs) measurements are not compatible with the α-ketol rearrangement mechanism. The hydroxyacetone and glycoaldehyde phosphate intermediates have not been directly observed. Neither have they been successfully incorporated when incubated with DXR and the necessary cofactors. These conflicting results indicate there is further work to be done on the DXR mechanism. Despite the rearrangement mechanism, the subsequent reduction produces MEP in the same way. Deuterium-labeled NADPH and crystal structures have revealed details of the reduction reaction. The Re face of MEsP protonated by the pro-S hydrogen from the nicotinamide C4 to the C1 of the aldehyde.

4-Diphosphocytidyl-2C-methyl-D-erythritol Synthase (CMS/IspD)

The discovery of DXS and DXR allowed for the identification and characterization of additional NMA enzymes in quick succession. Utilizing [2-14C]-labeled MEP in E. coli, research groups were able to track the formation of new radioactive products. The first product identified was 4-diphosphocytidyl-2C-methyl-D-erythritol (CDP-ME) in nuclear magnetic resonance (NMR) spectroscopic assays (Figure 1.4). A database search of corresponding enzymatic activity pointed researchers towards the ygbP gene. Subsequently, several experiments showed MEP turnover and IPP production are dependent upon ygbP.
and several other open reading frames\textsuperscript{101,155}. The distribution of the new protein correlated well with the expected presence of the NMA pathway in eubacteria. Following the confirmation of ygbP, now designated IspD (CMS), activity assays revealed the necessity of a divalent cation (Mn\textsuperscript{2+}, Mg\textsuperscript{2+}, or Co\textsuperscript{2+}) with a preference for Mg\textsuperscript{2+}. CMS appears to be substrate specific with low but measurable activity with GTP and ATP. The incorporation of CTP’s α-phosphate instead of the β- or γ-phosphates was confirmed utilizing radioactive phosphorous isotopes\textsuperscript{101,155}.

\[
\begin{align*}
\text{MEP} & \xrightarrow{\text{CTP, PP}_{\text{i}}} \text{CDP-ME} \\
\text{H}_3\text{C}-\text{OH} & \xrightarrow{\text{H}_3\text{C}-\text{OH}} \text{PO}_{\text{3}}^{\text{2-}} & \text{C}_{\text{3}}\text{H}_5\text{OH} & \xrightarrow{\text{OH}} \text{PO}_{\text{3}}^{\text{2-}} & \text{C}_{\text{3}}\text{H}_5\text{OH} & \xrightarrow{\text{OH}} \text{PO}_{\text{3}}^{\text{2-}} & \text{PO}_{\text{3}}^{\text{2-}} & \text{PO}_{\text{3}}^{\text{2-}} & \text{PO}_{\text{3}}^{\text{2-}}
\end{align*}
\]

Figure 1.4: The abridged representation of reaction catalyzed by CMS/IspD which attaches a cytidyl group to the phosphate tail of 1-deoxy-D-xylulose 5-phosphate producing 4-diphosphocytidyl-2C-methyl-D-erythritol.

Several crystal structures of CMS have been solved from a variety of organisms\textsuperscript{62,88,152}. To continue the trend of the first two enzymes, CMS is found commonly as a homodimer. Each structure revealed strong overall structural conservation with each other and other nucleoside-binding proteins, particularly cytidyltransferases\textsuperscript{152}. The domain of each monomer hold a certain characteristic inline with other homologues. One of the domains consists of a so-called β-arm, composed of overlapping parallel and anti-parallel β-strands, which protrudes from the main globular domain at a wide angle. This β-arm acts as a hook-like structure that interlocks closely with another monomer which aids in dimerization\textsuperscript{62,88,152}. The tertiary structures with all necessary cofactors provided valuable insights into ligand binding and enzymatic activity. A large network of hydrogen-bonding interactions between ligands and side chains as well as backbone carbonyl and amide groups revealed that both the substrate and products are fixed to the active site. Direct interaction between the protein and cytosine moiety of CTP, in part, explains the selectivity and preference for pyrimidines over purine nucleosides\textsuperscript{152}. Basic
residues are proposed to position the triphosphate tail for catalysis. Additionally, these residues might play a role in stabilizing the pentacoordinate transition state during the reaction. Additional phosphate coordinations of MEP and CTP is provided by the Mg^{2+} despite its lack of direct interactions with the enzyme^{62,88}.

The crystal structures of CMS has lead to the proposal of 2 reaction mechanisms. One proposal involves the formation of a reactive metaphosphate CMP molecule via elimination of a disphosphate group. The metaphosphate CMP is subsequently attacked by the 4-phosphate of MEP to form CDP-ME. The alternative mechanism proposed starts with a direct nucleophilic attack on the α-phosphate of CTP by the 4-phosphate of MEP. The collapse of the pentacoordinate intermediate produces CDP-ME and PPi. Current mutagenesis and structural data favor the second mechanism over the first^{152,154}.

4-Diphosphocytidyl-2C-methyl-D-erythritol Kinase (CMK/IspE)

The expanding knowledge of the first three enzymes continued to facilitate the discovery of the next downstream catalyst. Genomic analyses showed *E. Coli ychB* and its orthologous sequences showed similar patterns in eubacteria and plants as other NMA genes. Overexpression, purification, and characterization of *ychB* revealed the production of 4-diphosphocytidyl-2C-methyl-D-erythritol 2-phosphate (CDP-MEP) from CDP-ME (Figure 1.5); which corresponds to a phosphorylation of the C2 hydroxy group^{117}. Subsequently, the reaction and structure of ychB, later designated CMK or IspE, resembles those catalyzed by the GHMP superfamily of enzymes. In addition to galactokinases and homoserine kinases, two enzymes of the MVA pathway, mevalonate and phosphomevalonate kinases, defines the enzymes of GHMP superfamily^{117,185}.

Following from the observed sequential conservation, CMK homologues strongly resemble each other as well as other members of the GHMP superfamily. One distinction
between CMK and other GHMP enzymes, CMK is commonly found in a monomeric state where as GHMP family members are commonly found in a homodimeric structures\(^{85,126,168,185}\). Each CMK monomer consists of predominantly 2 domains. An N-terminal domain responsible for cofactor binding and a C-terminal domain in charge of substrate binding. CMK has an overall clamshell-like shape and the catalytic center is formed in an open cavity between the domains after the clamshell closes. This closure brings the substrate and cofactor into proximity in order to promote phosphorylation of CDP-ME\(^{126,168,169,185}\).

Based on similarities to GHMP kinases, a catalytic mechanism was proposed. Not on direct observations of the actions of the actual enzyme\(^ {61,97}\). The C2 hydroxyl group of CDP-ME forms hydrogen bonds with the side chains and carboxyl groups of highly conserved lysine and aspartate residues. This network helps to further polarize the hydroxyl group\(^ {126,168,185}\). Due to this polarization, one of the aspartate residues can act as a base to deprotonate the hydroxyl group. The resulting reactive alkoxide undergoes nucleotide attack of the $\gamma$-phosphate of ATP resulting in a similar pentacoordinate intermediate for CMS/IspD. The subsequent collapse of the intermediate results in ADP and CDP-MEP being released with turnover\(^ {126,168,185}\). A divalent metal ion is required for catalytic activity similar to other GHMP family members. The ion is responsible for positioning and orienting the phosphate moiety in proximity for attack by the acceptor molecule. Additionally, it stabilizes the pentavalent transition state the bond between the $\beta$- and
γ-phosphates of ATP\textsuperscript{34,61,70,97}. Though, no CMK crystal structure has been observed to contain the metal ion and lack of a highly conserved glutamate residue involved in positioning the metal ion suggests unique role in CMK. Additionally, some have proposed coordinated water molecules might act in place of the metal ion in certain kinases. The exact role of the metal remains to be determined\textsuperscript{34,126,168,185}.

2C-Methyl-D-erythritol-2,4-cyclodiphosphate Synthase (MCS/IspF)

When CMS was identified as the third enzyme of the NMA pathway, the \textit{ygbP} gene expression was found coupled to another unannotated \textit{ygbB} gene sequence with a few cases even fused inside a single open reading frame\textsuperscript{75,155}. Correspondingly, the species distributions of the gene orthologues parallel the presence of NMA based isoprenoid biosynthesis. Attempts to identify the activity of the corresponding protein was determined by challenging the enzyme with CDP-ME and CDP-MEP which produced 2-C-methyl-D-erythritol-3,4-cyclophosphate (MEcP) and 2-C-methyl-D-erythritol-2,4-cyclodiphosphate (MEcDP), respectively\textsuperscript{75,181}. MEcDP was found to have been previously detected as a bacterial metabolite. These results support MEcDP as a new intermediate between DXP and IDP or DMADP (Figure 1.6); while MEcP was regarded as an in vitro artifact with no physiological relevance. Subsequently, the name of the enzyme was changed MEcDP synthase or IspF to reflect its function and position in the pathway\textsuperscript{75}.

\begin{figure}[h]
\centering
\includegraphics[width=0.8\textwidth]{figure1_6.png}
\caption{The cyclization of 4-diphosphocytidyl-2C-methyl-D-erythritol 2-phosphate and subsequent expulsion of cytidyl release catalyzed by MCS/IspF.}
\end{figure}
Structural characterizations have been published for the E. coli, Plasmodium falciparum, Plasmodium vivax, and A. thaliana MCS enzymes\textsuperscript{28,87,139,153,177}. While there are differences in the spatial grouping and composition of the asymmetric unit, the structures all showed the formation of a tightly associated homotrimer. The multimeric assembly buries a large surface area that contributes to the MCS enzyme’s heightened stability even in the face of mass spectrometric analysis. This trimeric structural feature is a common feature observed between MCS and any of its wider structural or functional homologues\textsuperscript{87,153}. A series of anti-parallel β-sheets form a largely hydrophobic channel at the core of the trimer. This channel is thought to play a role in feedback regulation\textsuperscript{87,89,153,177}. The active site is found at the interface of two subunits with both side chains contributing to the catalytic center. Conformational stabilization of the substrate and intermediates is accomplished via interaction with a few key amino acids, and two distinct essential metals, a Zn\textsuperscript{2+} and either a Mg\textsuperscript{2+} or Mn\textsuperscript{2+}. A zinc ion responsible for positioning the cytidyl moiety of the substrate; which itself is tetrahedrally coordinated by an aspartate and two histidine residues as well as the β-phosphate of MEcDP. Both the α- and β-phosphates of the CDP substructure is coordinated and stabilized by either a Mg\textsuperscript{2+} or Mn\textsuperscript{2+}. These phosphate groups additionally play a role in the octahedral coordination of the Mg\textsuperscript{2+} or Mn\textsuperscript{2+} ions with three water molecules and a glutamate residue filling in the rest of the coordination sites\textsuperscript{87,153,177}.

The intramolecular cyclization of CDP-MEP to MEcDP and concomitant CMP release catalyzed by MCS is thought to proceed via an in-line mechanism. Analogous to the previous two enzymes, the reaction involves the nucleophilic attack on a phosphate moiety thus forming a pentacoordinated transition state. The subsequent collapse of the transition state releases the two products, CMP and MEcDP\textsuperscript{87,153,177}. The protective flexible loop closes off the catalytic cavity from the surrounding solvent. Interactions with CDP and MEP substructures via hydrogen-bonding and hydrophobic regions of the cavity accountants for the high degree of selectivity of MCS. The diphosphocytidyl moiety align-
ment is accomplished primarily by the active site metal ions with additional help from hydrogen-bonding and hydrophobic interactions of active site residues. Of particular interest for reactivity, the Zn$^{2+}$ ion increases the electrophilic character of the $\beta$-phosphate in addition to aiding in lining up the nucleophilic attack by the 2-phosphate of the MEP moiety. Additionally, the enzyme restricts the flexibility of the substrate bringing the electron donor and acceptor in close proximity$^{153}$. The negative charge of the cyclic transition state is compensated by the positive charge of the 2 metal ions$^{87,153,177}$.

1-Hydroxy-2-methyl-2-(E)-butenyl 4-diphosphate Synthase (HDS/IspG)

Unlike the previous upstream catalysts were discovered and characterized in relatively quick succession, the final two steps proved more elusive. The unannotated $gcpE$ gene was originally discovered in association with a histidyl tRNA synthetase$^{59}$. Utilizing bioinformatic approaches, an association was observed between the $gcpE$ gene and other NMA pathway enzymes were observed to reflect the characteristic distribution patterns. As with DXS, DXR and IspD-IspF, the new gene sequence was detected in various bacterial species, plants and apicomplexa, but not in eukaryotes such as yeast$^{2,29}$. Disruption of this gene was lethal as with the other NMA genes. Isotopic labeling coupled with NMR analysis identified the new intermediate as 1-hydroxy-2-methyl-2-(E)-butenyl 4-phosphate (HMBDP) (Figure 1.7), which can be produced via a reductive deoxygenation of MEcDP$^{71}$. The gene was renamed to IspG to reflect the new position in the NMA pathway. Subsequent, recombinant expression and purification was straightforward, observed activities were low. The presence of three highly conserved cysteines and similarities with sequence motifs of ferrodoxin and aconitase enzymes suggested the presence of a catalytic iron-sulfur, [4Fe4S]$^{71,192}$. Further assays performed under anaerobic conditions and the presence of an reducing agent for regenerative purposes resulted in the efficient production of HMBDP from MEcDP. Additional confirmation of the presence of the cluster was UV-vis absorption spectrum, which matched previously observed spectrum of simi-
As with DXS, DXR and CMS, HDS is commonly found in a homodimer. Each monomer is composed of two domains. The N-terminal domain is an 8-stranded β-barrel globular subunit similar to the common (βα)8-fold of the triose phosphate isomerase (TIM) barrel superfamily. The iron-sulfur cluster is found in the C-terminal domain. Coordination of the cluster is supplied via 3 cysteine residues and a glutamate residue. MEcDP binds in the active formed between the C-domain of a monomer and the N-domain of the other monomer. All published HDS crystal structures are highly similar; particularly with respect to the N-domain. The C-domains are absent in some structures (probably due to lack of resolution), while the *Plasmodium falciparum* structure has an additional domain. This additional domain is thought to fold into a second TIM barrel to allow for monomeric activity.

Mechanistic details remained elusive despite the identification of the substrate, product and resolution of several crystal structures. The reaction was known to involve the [4Fe4S] cluster, elimination of the C3 hydroxyl and a 2 electron reduction. Results of isotopic-exchange experiments, electron paramagnetic resonance (EPR) spectroscopy, and many other experiments permitted the description of the HDS mechanism. Upon binding MEcDP, a conformational closure causes the displacement of the glutamate residue from the fourth iron while promoting the formation of a covalent bond with the substrate. A deprotonation of the C3 hydroxyl group by a
second glutamate assists in the Fe-O bond formation\textsuperscript{106,150,204}. Once bond, the ring of MEcDP opens and closes consistently\textsuperscript{198}. The introduction of the first single external electron breaks the ring permanently either a carbocation or radical (formed via internal electron transfer) and begins the reaction in earnest. The addition of a second external electron produces a C2 carbanion\textsuperscript{146,186,187}. Formation and release of HMBDP proceeds via an E\textsubscript{1cb} elimination results. A localized proton relay change results in the release of H\textsubscript{2}O from the cluster and regeneration of the enzyme\textsuperscript{146}.

### 1.3 Isoprenoids as Drug Target

Molecular medicine has provided means for mankind to overcome many diseases caused by various microbial life forms. Lately, there has been a growing resistance to current therapies. The discovery of multi-drug resistance forms of many diseases (e.g. tuberculosis and malaria) are poised to return us to the time prior to anti-microbial drugs\textsuperscript{96,122,165}. In an age of growing drug resistance, there are very few companies investing in developing novel treatments due to the low returns and high upfront costs. Amongst the now growing list of neglected disease, Tropical diseases (i.e. malaria, leishmaniasis, tuberculosis) represented the most neglected diseases in the world. This due in large part to their concentration in the developing nations. Malaria is one of the most profound problems due to its high morbidity rate and millions of reported cases a year. In the age of modern globalization, growing resistance is a world wide problem\textsuperscript{122,183}.

Malaria presence a growing international concern. High mortality aside, malaria can cause economic downturns in high areas of infection due to an acutely infected individuals inability to work. Long term effects can be seen by life-long learning impairments caused when children are infected. Malaria is caused by four species of \textit{Plasmodium} but the majority of the mortalities are cause by two of them, \textit{P. falciparum} and \textit{P. vivax}\textsuperscript{24,183}. Both show evidence of a growing resistance to long standing therapies such as chloroquine and
fansidar; which has hastened the need to develop novel treatments.

Primarily, drug resistance has been found via mutations in enzymes which reduce the inhibitive effects of the therapy. Additional mutations found in transporter proteins make up a large portion of the remaining resistance\textsuperscript{196}. These transporters (i.e. \textit{pfmdr1}) act by removing the drug from the target sites. This is similar to the developing of $\beta$-lactamases in infectious bacteria to destroy antibiotics such as penicillin while developing mutations in the target peptidase enzymes. It is a two fold development in resistance.

The NMA pathway has great promise as a target for anti-microbial activity. The most significant benefit is the seeming absence of orthologous enzymes in mammalian cells. Particular absence in humans is a huge benefit. In other words, the entire pathway seems to be absent in humans which rely on the MVA pathway for IDP and DMADP production. The combination of the completion of the human genome project, subsequent expansion in mapping other species genomes, and the identification of the genes and enzymes of the NMA pathway allowed researchers to perform scans. Table 1.2 represents highlights of species and the isoprenoids biosynthesis pathways present. There are a few species that rely on both but have one isolated in an organelle, therefore the products of one of the pathways aren’t readily available to use in a crisis\textsuperscript{59,63}.

As indicated in table 1.2, several protozoal genomes (e.g. \textit{P. falciparum}, and \textit{P. vivax}) have genes corresponding to the NMA pathway. Subsequent studies have revealed these genes to be predominantly located in the apicoplast; which is necessary for survival in the intraerythrocytic and intrahepatic stages of \textit{plasmodium}. The inhibition of the NMA pathway via fosmidomycin can only be save via exogenous introduction of IDP and DMADP suggests this pathway as a new source of anti-malarial drugs\textsuperscript{59,201}. 
Additionally, there is evidence of NMA being a good source of anti-bacterial drugs. A majority of current antibiotics work via the interruption of the biosynthesis of macromolecular components (e.g. DNA, RNA, cell wall) of the bacterium$^{6,52}$. The remaining
<table>
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Table 1.2: Highlights the existence of genes found for both the Mevalonate and Non-Mevalonate Pathways in a variety of life forms.
few target metabolic enzymes. Arigoni et al. conducted a study using bioinformatics to identify 30 *E. coli* necessary for survival which could also be found in other bacterial species. The NMA pathway is amongst these necessary genes\(^6\). Several pathogenic bacteria, including *E. coli* and *Mycobacterium tuberculosis*, carrying deletions in the NMA genes could only be rescued via exogenous introduction of isoprenoids\(^55,122\). The presence of the NMA pathway in several pathogenic species but absence in ours indicates this pathway as a key source of novel therapies to combat the growing resistance problem. There are a couple of issues. There are two enzymes belonging to large families of enzymes. DXS and IspE belong the TDP-dependent enzyme and GHMP families, respectively. High sequence similarity between these enzymes and their families poses unintended consequences. This has been observed consistently in work with kinases. Trying to develop a highly specific inhibitor is troubling at best. These considerations should not inhibit our attempts at developing new therapies base on this pathway.

### 1.4 Computational Methodology

Biochemistry is the study of the chemical reactions involved in biological processes. At the heart of this endeavor are enzymes that facilitate these processes. Hence, it became rapidly apparent a deeper understanding of enzymes was needed. When studying enzymes some key questions are: “What amino acids are participating in the enzymatic action? What are their roles? And what are the possible transition states?”\(^116,148,182\). In pursuit of answers to these questions, biochemists developed laboratory techniques to probe the relative importance of certain amino acids (AAs) and the motions of these highly dynamic macromolecules. Some of these experimental methods are kinetic isotope effects (KIE); site directed mutagenesis, and Forster resonance energy transfer (FRET). It was hoped that they could give insights into transition state structures and a better view of the molecular level interactions occurring in enzyme active sites. These techniques have contributed significant knowledge of the inner workings of the enzymes, however
they do have limitations. With the advent of macromolecular crystallography and later NMR based methods, the ability to see at the molecular level was greatly enhanced.

Computational biochemistry can provide an even more detailed look into enzyme mechanisms. This includes but is not limited to the investigation of the motion of enzymes, de Novo design of transition state analog inhibitors, and investigating protein-protein interactions\textsuperscript{93}. My current focus is in 3 areas: application of computational techniques to elucidate mechanistic detail of two enzymatic processes.

The study of condensed phase chemical and biochemical processes has been major focus for both experimental and computational chemists for several decades now. Though QM approaches for computation are more accurate, the computational cost prohibits the use of these approaches with any biologically relevant systems. This limitation of QM methods was a driving force behind the growing use of more efficient MM methods that are more empirically driven. Significant time and effort has been put into attempting to improve the efficiency of traditional QM methods recently. Though these advances have shown benefits for small molecule chemistry, they are still prohibitively expensive for full scale biochemical applications. A problem that has been mitigated in part through the development of more efficient QM codes and the growing use of hybrid QM/MM methods. Standard methods of QM/MM attempt to couple the cost efficiency of MM methods with the accuracy and precision of QM methods through the division of the system into subsystems. These subsystems are treated at different levels of theory. One region, that is usually made up of the active site or site of most significant interest, is labelled the QM region and treated with the highest level of computational theory. An MM region is also defined and the protein environment that surrounds the QM region. The third region is an interface region that connects the QM and MM regions previously defined. The third region is only necessary if in the course of defining the QM region from the MM region, any bonds found along their borders are split between the regions, becoming the interface.
region. A coupled potential is responsible for the inclusion of electrostatic and van der Waals interactions from the QM and MM through the interface region\textsuperscript{136,182,188}.

Several methods for the implementation of hybrid QM/MM schemes have been reported. Empirical valence bond (EVB) and semi-empirical methods have been employed typically to describe the QM region, due to their relative efficiency in comparison to ab initio QM theory. Though they have been used effectively, several weaknesses that have been well documented. Recently, there has been a bigger push to overcome the deficiencies in these methods through the implementation of more accurate and rigorous computational methods such as ab initio and Density Functional Theory (DFT). Herein, we will be applying QM/MM methodology with the QM region being treated with the more rigorous DFT methodology\textsuperscript{162}.

A major advantage of using hybrid QM/MM techniques is the ability to compute barriers for biological processes (e.g. NOX production). The relative free energy ($\Delta G$) of each step along the reaction will be calculated in order to ascertain (within relative degrees of certainty) the profile of a proposed mechanism and therefore suggest the most energetically favorable mechanism\textsuperscript{136,182,188}. $\Delta G$ has been defined as a measure of the driving force behind a reaction. Thus by calculating the driving force, the mostly probable reaction will be uncovered. In addition, key residues involved in the stabilization of the transition state or destabilization of the reactant state will be identified and analyzed for relative electrostatics in determining the relative energetics of different reaction mechanisms.
Chapter 2

Thiamin Diphosphate Activation in 1-deoxy-D-xylulose 5-Phosphate Synthase: Insights into the Mechanism and Underlying Intermolecular Interactions

2.1 ACS Permissions


2.2 Abstract

1-deoxy-D-xylulose 5-phosphate synthase (DXS) is a thiamin diphosphate (TDP) dependent enzyme that marks the beginning of the methylerythritol 4-phosphate isoprenoid biosynthesis pathway. The mechanism of action for DXS is still poorly understood and begins with the formation of a thiazolium ylide. This TDP activation step is thought to proceed through an intramolecular deprotonation by the 4’-aminopyrimidine ring of TDP; however, this step would occur only after an initial deprotonation of its own 4’-amino group. The mechanism of the initial deprotonation has been hypothesized, by analogy to transketolases, to occur via a histidine or an active site water molecule. Results from hybrid quantum mechanical / molecular mechanical (QM/MM) reaction path calculations
reveal an ∼10 kcal/mol difference in transition state energies, favoring a water mediated mechanism over direct deprotonation by histidine. This difference was determined to be largely governed by electrostatic changes induced by conformational variations in the active site. Additionally, mutagenesis studies reveal DXS to be an evolutionarily resilient enzyme. Particularly, we hypothesize that residues H82 and H304 may act in a compensatory fashion if the other is lost due to mutation. Further, nucleus-independent chemical shifts (NICS) and aromatic stabilization energy (ASE) calculations suggest that reduction in TDP aromaticity also serves as a factor for regulating ylide formation and controlling reactivity.

2.3 Introduction

Isoprenoids are one of the largest and most diverse families of biomolecules with a number of them essential for life. An example would be Vitamin A, which plays a role in human growth and development as well as immune system maintenance. Two isoprene molecules are variably employed in the construction of all isoprenoids. Isopentenyl diphosphate (IDP) and dimethylallyl diphosphate (DMADP) are produced via two distinct biosynthetic pathways (Figure 2.1): mevalonate pathway (MVA) and methyler-thritol 4-phosphate pathway (MEP pathway). The MVA pathway was discovered in the 1950s and was considered the sole pathway until the 1990s when discrepancies in isotopic labeling studies led researchers to hypothesize an alternative, MEP pathway. Subsequent, genetic studies have revealed a large variety of life (e.g., algae, bacteria, etc.) to have varying degrees of dependence upon MEP pathway for isoprenoid production; in addition to a number of human pathogens (e.g., Plasmodium spp., and M. tuberculosis). Interestingly, the MEP pathway is absent in all mammalian genomes meaning that the enzymes of this pathway are ideal targets for novel antibiotics and antimalarials. For example, fosmidomycin is known to be an effective agent against several of the Plasmodium spp. (malarial pathogens) and targets MEP pathway’s second step, 1-deoxy-D-xylulose 5-phosphate reductoisomerase (DXR).
Figure 2.1: Schematic of isoprene production via MVA or MEP pathway. MVA pathway produces DMADP via a secondary enzyme, IDP Isomerase\(^{45,171}\). MEP pathway directly synthesizes both isoprene molecules.

MEP pathway is comprised of eight reactions catalyzed by seven enzymes beginning with 1-deoxy-D-xylulose 5-phosphate synthase (DXS)\(^{18,53,69,103,114,115,124,176}\). DXS catalyzes the condensation of glyceraldehyde-3-phosphate (G3P) and pyruvate to produce 1-deoxy-D-xylulose 5-phosphate (DXP). Aside from isoprenoid production, DXP is utilized in the production of vitamin B\(_1\) (thiamin) and vitamin B\(_6\) (pyridoxine) biosynthetic pathways\(^{14,53,76}\) suggesting increased significance to understanding the mechanism of DXS. Further, DXS is believed to be a rate-limiting step due to the observed correlation between isoprenoid product levels and DXS levels.\(^{53}\) DXS is a member of the thiamin diphosphate (TDP) dependent family of proteins; specifically a member of the transketolase (TK) enzyme subclass and also possesses pyruvate decarboxylase activity.\(^{7,114,176}\) TKs are a class of TDP dependent enzymes responsible for the transfer of a ketol donor group to an aldehyde or ketone acceptor molecule.\(^{41,60,163}\) In 2007, Xiang et al. published crystal structures and mutagenesis results of DXS from \textit{E. coli} and \textit{D. radiodurans} and compared them to the E1 subunit of pyruvate dehydrogenase (PDH) and yeast TK (members of the same class of enzymes).\(^{197}\) The comparison revealed significant similarities between these four enzymes: 1) each enzyme is composed of three domains (I, II, and III), 2) all possess a TDP cofactor, and 3) all contain a GDGX\(_{25-30}\)N motif that
plays a role in producing the twisted ‘V’ shape of the TDP cofactor in the active site\textsuperscript{197}. The strained cofactor conformation has been shown to play a role in lowering the pK\textsubscript{a} of a hydrogen on the thiazolium ring’s C2 atom (Figure 2.2)\textsuperscript{25,26,60,84,125}. The active site of DXS contains a number of strictly/highly conserved residues (e.g., Glu370/372, Asp152/154 in \textit{E. coli}/\textit{D. radiodurans}, respectively) that are common among TDP dependent enzymes, particularly TKs. These similarities have led researchers to propose a DXS reaction mechanism based, primarily, on mechanistic data of other TK enzymes (Figure 2.2)\textsuperscript{41,163,173}.

![Proposed general mechanism for DXP biosynthesis](image)

Figure 2.2: Proposed general mechanism for DXP biosynthesis. Pieces of each step are labeled with different colors to indicate where they originate from. Red represents pyruvate and blue are the pieces affiliated with G3P.

Although DXS contains many of the strictly/highly conserved residues of the TDP-dependent superfamily of enzymes (\textit{vide supra}), it displays distinct structural features\textsuperscript{197}. Specifically, the domain arrangement of DXS; homodimeric with a deep pocket between
two domains of the same monomer rather than at the dimer interface as is the case with other TDP enzymes. These structural differences logically lead to questions of mechanistic similarity. Until the discovery of DXS, it was believed that all TDP enzymes functioned via a classical “ping-pong” mechanism (i.e., pyruvate binding $\rightarrow$ CO$_2$ release $\rightarrow$ G3P binding)\textsuperscript{60}. However, Eubanks and Poulter in 2003 concluded that DXS operates via an ordered mechanism (i.e., irreversible pyruvate binding $\rightarrow$ G3P binding $\rightarrow$ CO$_2$ release) and hypothesized a side reaction for producing CO$_2$ via binding of a second pyruvate molecule\textsuperscript{54}. This hypothesized side reaction was subsequently confirmed by Brammer and Meyers in 2009\textsuperscript{19}. However, the following year a steady-state kinetics study examining a herbicide metabolite (ketoclomazone, a derivative of clomazone) provided evidence of a traditional ping-pong mechanism for DXS\textsuperscript{123}. Nearly, simultaneously (in 2010), a single-molecule force spectroscopy nano sensor was developed and used to observe an approximate 2-fold binding enhancement of G3P in the presence of pyruvate; suggesting an ordered DXS mechanism\textsuperscript{174}. As part of this work, the authors cast doubt on the reliability of previous results based upon assays that measure bulk phenomena rather than single-molecule behavior. To further confound the situation, Meyers and co-workers (in 2011) proposed an unprecedented TDP-based mechanism; G3P and pyruvate were found to bind independently and reversibly. Thus, they concluded DXS functions via a rapid equilibrium, random sequential mechanism\textsuperscript{20}. In the following year, Meyers and co-workers revealed a 600 fold acceleration in the decarboxylation of the lactyl-TDP intermediate upon binding of G3P. This result further distinguishes DXS from other TDP-dependent enzymes.

Two recent studies have called into question our understanding of the active sites of the large class of TDP dependent enzymes. For instance, the benzaldehyde lyase (BAL) enzyme is devoid of all but two acid-base residues around the TDP active site: a histidine and highly conserved glutamate\textsuperscript{22,121}. Most interesting is the lack of any apparent acid-base residues in glyoxylate carboligase (GCL)\textsuperscript{86}. These recent discoveries represent glaring gaps in our understanding of TDP-dependent enzymes and bolster the importance
of investigating distinct related enzymes (i.e., DXS).\textsuperscript{21,22,86,121} Here, computation is an ideal partner to experiment.

Of particular interest in this work is the “true first step” of this process: TDP activation, of which significant mechanistic details are still largely uncertain. For example, a proposed mechanism for ylide formation begins with an initial deprotonation of the 4′-aminopyrimidine (AP) state that produces the 1′,4′-iminopyrimidine (IP) state\textsuperscript{114,176}. A general base (GB) is required for this deprotonation, however, the identity of this group remains unknown. One hypothesized GB is a highly conserved histidine (His434 in \textit{D. radiodurans} DXS) found proximal to TDP’s 4′-amino group\textsuperscript{60,82}. The aforementioned mutation studies (i.e., H434A) showed approximately 95% activity retention, which suggests an alternative mechanism. A more recent 2014 study by Querol et al. suggests H431 (\textit{E. coli} equivalent of \textit{D. radiodurans} H434) plays a role in transition state stabilization but not required for catalysis\textsuperscript{145}. Additionally, numerous structural differences between DXS and TK enzymes (\textit{vide supra}) support the possibility of an alternative mechanism\textsuperscript{197}. One possible alternative mirrors that of human TKs; where a water molecule would replace H434 as the GB with a Gln residue acting to stabilize this via coordination\textsuperscript{134,173,190}. This results in two possible TDP activation mechanisms: a water-mediated mechanism (WMM) or direct histidine mechanism (DHM). Even though the WMM utilizes a water molecule as the initial general base, it is possible that H434 plays a role in this mechanism as either a coordination site for the water molecule or as the final location of the proton.

TDP has been shown to exist in four different tautomeric/ionization states (Figure 2.3); however, the exact mechanism for producing the final ylide form remains unclear\textsuperscript{8,9,133,141}. Figure 2.3 illustrates two possible mechanisms: (1) a concerted AP to IP conversion followed by ylide formation or (2) a step wise mechanism where the AP is first ionized to a 4′-aminopyrimidinium ion (APH\textsuperscript{+}) and, subsequently, converted to the IP and ylide, respectively. Although direct spectroscopic evidence of the APH\textsuperscript{+} state remains elusive, its existence has been inferred from alternative experiments (e.g., pH rate profiles, solid-state NMR) and hypothesized to assist in promoting IP formation via
stabilization of the tautomerization reaction. An elevation in the pKₐ of TDP’s N1 atom is proposed to account for the APH⁺ state’s existence; which is justified by its proximity to a strictly conserved glutamate residue. This idea, however, does not consider the possibility of an accompanying elevation in the pKₐ of the glutamate residue.

Recent studies have determined the pKₐ for the N1 atom in DXS to be 7.5 and a PROPKA calculation has estimated the E373 residue to have a pKₐ of 8.4. These pKₐ values suggest an equilibrium between the AP, and APH⁺ states; which is consistent with the enzyme stabilizing the IP formation via pKₐ modulation. Additionally, Jordan et al. supports the equilibria presented in Figure 2.3; particularly for apo (TDP-bound enzyme lacking substrate) enzymes. Therefore, it is not necessary to select between the step wise or concerted mechanism for the purposes of this study.

Figure 2.3: Structure and relationship of the 4 possible tautomeric/ionization states proposed for the cofactor of TDP dependent enzymes. Key atoms have been given names for reference purposes throughout this article.

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1 The pKₐ for the N1 atom of DXS was determined via pH rate profile studies. The E373 pKₐ was estimated using PROPKA3.1 with TDP but without pyruvate and G3P. The complete output of the PROPKA3.1 calculation can be found in the supporting information.
The rate of activation and turnover of TDP shows a substantial increase when bound to an enzyme rather than in solution\textsuperscript{84}. Several factors leading to the increase in activity have been proposed. One factor includes the strained conformation the cofactor adopts upon binding. This conformation places the 4′-amino group in close proximity to the thiazolium C2 atom; both introducing strain and lowering the pK\textsubscript{a} of the C2 hydrogen from 14-19 (depending on solvent) to approximately 9\textsuperscript{65,82}. The electronics of the pyrimidine ring of TDP would also undoubtedly change during the activation process. These changes could lead to a disruption of the aromaticity and more indirectly influence the energetics of TDP activation. The link between aromaticity and TDP activation has not been investigated previously. Uncovering such a link will provide better understanding of DXS and raise the question if this phenomenon is general for all TDP dependent enzymes (e.g., TK, PDH, etc.).

Herein, a hybrid quantum mechanical/molecular mechanical (QM/MM) study is carried out using the \textit{D. radiodurans} DXS crystal structure\textsuperscript{197}. The energy profile of the DHM and WMM are computed to determine the most favorable activation mechanism. Active site electrostatics are also probed to elucidate the stabilizing/destabilizing effects that govern this process. Further, two metrics of aromaticity are employed to quantify this effect and determine its role as a possible driving force in activation of TDP dependent enzymes. In addition to the computational work, a kinetics study, utilizing a coupled enzyme assay, was performed on mutant and wild-type forms of the \textit{D. radiodurans} DXS enzyme. As part of the mutagenesis work, we have re-examined the H434A mutation; which is of particular interest in this study. Our study focuses on the K\textsubscript{M} and k\textsubscript{cat} due to their relationship with substrate affinity and reaction turnover, respectively. The combination of computational and experimental results helps bridge the gap between bulk behavior and atomistic understanding; ultimately leading to new insights into this unique enzyme.
2.4 Methods

2.4.1 Computational Methods

The DXS active homodimer structure was used throughout this study. The crystal structure for the *D. radiodurans* DXS (PDB ID:2O1X)\textsuperscript{197} enzyme with TDP bound was processed and parsed via www.charmming.org\textsuperscript{127}. A TDP molecule can be broken down into three moieties: a thymine-like pyrimidine ring, a pyrophosphate (residue name utilized in the topology file), and a thiazolium ring(Figure 2.2). Parameters for all three of these have been developed as part of the CHARMM General Force Field (CGenFF)\textsuperscript{184}. Final TDP parameters were thus obtained by connecting the respective components and modifying charges (see SI). Parameter validation was done with respect to the TDP crystal conformation based on the RMSD (see SI).

Structural modifications were performed to ensure the active site Glu373 was protonated in agreement with experimental evidence\textsuperscript{90,112}. CGenFF and CHARMM22 protein (C22) force fields\textsuperscript{119} were used throughout. The system was solvated in a rhombo-dechedron crystal structure and neutralized with KCl salt to a final concentration of 0.15M. The system was heated from 110K to 310K over 100ps and equilibrated for 200ps at constant pressure (1atm) and temperature (310K). The total system size was then reduced by removing all of the water and salt ions beyond 12Å from the surface of the protein. The reduced structure was then QM/MM minimized, without applying cutoffs, to a gradient tolerance of 0.002 kcal·mol$^{-1}$·Å$^{-1}$. All QM/MM calculations employed the Q-Chem4.0\textbackslash CHARMM\textsuperscript{27,170,194} interface at the B3LYP/6-31G* level of theory.\textsuperscript{13,68,105} Additionally, the single link atom scheme was used to account for truncation of the QM region and employed group electrostatic exclusions to prevent over polarization of the QM region.

Reaction pathway calculations employed a combination of the replica path method (RPATh) and harmonic distance restraints (RESDi)\textsuperscript{36,194,195}. The RPATh method permits the user to divide the system into discrete subsystems (replicas), which are allowed
to change independently of the remaining “environment”. A subsystem was defined to be 6.5Å around the QM region, which was comprised of TDP, Glu373, His120, and His434 (Figure 2.4a, 98 QM atoms). The QM region for the WMM included a water molecule (Wat9709) coordinating with the 4’-amino group of the pyrimidine ring of TDP and the Nε of His434. Wat9709 was removed from the initial structure prior to QM/MM minimization (*vide supra*) for the DHM. Two replicas of the subsystem were used to model successive steps along the reaction coordinate (δ), which was defined as a linear combination of the bond being broken and the bond being formed (Figure 2.4b, 101 QM atoms). The δ values were defined incrementally for each mechanism starting from the reactant state. The DHM scanned a range of -3.0Å to 3.0Å in increments of 0.3Å with smaller increments of 0.1Å used around the transition state (i.e., -1.0Å to 1.0Å) to provide finer detail. The WMM scanned a range of -1.1Å to 1.1Å in increments of 0.1Å.

The Charge Perturbation Analysis (CPA)\textsuperscript{12,42,67,107} technique involves QM/MM single point energy calculations where a single residue’s classical charge is scaled to zero to probe its electrostatic contribution. ∆E is computed by taking the difference of the modified (zero-charge residue) and the full QM/MM electronic energy: \n
\[ \Delta E_{\text{CPA}} = E_{\text{elect}}^{\text{ZeroChargeRes}} (\text{QM/MM}) - E_{\text{elect}}^{\text{FullMM}} (\text{QM/MM}) \]

CPA calculations were performed for the reactant state (RS) and transition state (TS) of the DHM and WMM as determined by RPATh+RESDi calculations. The reactant state was the starting δ value for each mechanism while the TS corresponded to the point along the path with the highest energy. \n
\[ \Delta \Delta E_{\text{CPA}} = \Delta E_{\text{CPA}} (\text{RS}) - \Delta E_{\text{CPA}} (\text{TS}) \]

provides insight into stabilizing/destabilizing electrostatic effects with respect to RS and TS. CPA was performed on all 82 residues found within 5Å of the QM region for both mechanisms. To further characterize long range electrostatic changes, the QM/MM dipole moments were calculated around the QM region for the RS and TS of each mechanism. The QM/MM dipoles take into account the external charge contributions of the MM region on the QM region. The calculations were carried out using Q-Chem 4.0\textsuperscript{170} and initially visualized in IQmol with final rendering using PyMOL.
Two metrics were employed to quantify aromaticity and gauge the level of significance of it as a possible driving force of ylide formation: nucleus-independent chemical shifts (NICS)\textsuperscript{164} and aromatic stabilization energy (ASE)\textsuperscript{43,44}. NICS directly measures the aromatic character of a compound\textsuperscript{35,39,43,164} while ASE reveals the stabilization/destabilization that arises from the aromaticity of a compound\textsuperscript{1}. Due to computational limitations, NICS calculations were performed on reduced versions of the RS and TS subsystems. The reduced subsystems contained 232 or 235 atoms for the DHM or WMM, respectively. The difference of 3 atoms being the absence of Wat9709 from the DHM. Ghost atoms were placed perpendicular to the plane of TDP’s pyrimidine ring. Due to the non-symmetric protein active site, the NICS(0) (ring center), NICS(1), and NICS(-1) (atoms 1 Å above and below the plane of the ring) will be reported herein\textsuperscript{10,35,44,98}. All NICS calculations were performed using Q-Chem 4.0\textsuperscript{170} at the B3LYP/6-31G* level of theory\textsuperscript{13,68,105}. ASE is typically computed via a reference homodesmotic reaction\textsuperscript{1,189}. A homodesmotic reaction must be defined such that equal numbers of each type of atom (sp\textsuperscript{3}, sp\textsuperscript{2}, sp) and bond (sp\textsuperscript{3}-sp\textsuperscript{3}, sp\textsuperscript{3}-sp\textsuperscript{2}, sp\textsuperscript{2}-sp\textsuperscript{2}, etc...) exist in both reactants and products\textsuperscript{189}. All structures used in ASE calculations were optimized at the B3LYP/6-311+G** level of theory. Energies for each molecule were corrected by subtracting out their respective zero point energy obtained from subsequent frequency calculations.

2.4.2 Experimental Methods

Materials

TDP, pyruvate, G3P, DXP sodium salt, bovine serum albumin, and LB-broth were purchased from Sigma Aldrich. NADPH was purchased from Alexis Biochemical, Ni-NTA resin was purchased from Invitrogen, and β-mercaptoethanol (β-Me) was purchased from Fisher. \textit{E. coli} XL-10 cells, deoxynucleotide mix PCR grade, \textit{pfu} Ultra Hotstart DNA polymerase, QuikChange II site directed mutagenesis kit and acetonitrile (HPLC grade) were purchased from Agilent. The DNA vectors pET28a(+) and pET15b(+) and \textit{E. coli} BL-21 B(DE3) cells were purchased from EMD Biosciences. DNA sequencing services and
primers were purchased from MWG operon. All the other reagents were of the highest quality commercially available.

**Cloning of *D. radiodurans* DXS and *E. coli* DXR**

A synthetic, codon optimized *D. radiodurans dxs* gene with 5'-NdeI and 3'-XhoI restriction sites in a pMK vector was purchased from Geneart (Germany). The *dxs* gene was excised from the pMK vector and cloned into the *NdeI* and *XhoI* sites of a pET28a(+) vector (*kanamycin* resistance) with an N-terminal His$_6$-tag to yield the pET28a(+)-DXS plasmid. Successful cloning of the *D. radiodurans dxs* gene was confirmed by DNA sequencing at MWG Operon.

A synthetic, codon optimized *E. coli dxr* gene with 5'-NdeI and 3'-BamHI restriction sites in a pMK vector was purchased from Geneart (Germany). The *dxr* gene was excised from pMK vector and cloned into *NdeI* and *BamHI* restriction sites of pET15b(+) vector with a C-terminal His$_6$ tag to yield the pET15b(+)-DXR plasmid. Gene insertion was confirmed by DNA sequencing.

**Production of the *D. radiodurans* DXS Mutants**

Site-directed mutagenesis was carried out using the QuikChange II site-directed mutagenesis kit. Briefly, the mutagenesis mixture consists of 50-100 ng plasmid pET28a(+)DXS as a template, 1X PCR reaction buffer, 0.4 mM each of the forward and reverse primer, 0.25 mM dNTP mixture, 5 µL Quik solution, and 2.5 units of pfuUltra hotstart polymerase in a 50 µL reaction. The overlap extension method was used to produce the DXS mutants that were difficult to create via site directed mutagenesis$^{77}$. The sequence of the mutant DNA was confirmed by DNA sequencing.

**Assays for DXS Activity**

We employed a DXS-DXR coupled assay to determine the wild-type and mutant DXS enzyme activities. In this way, the DXS-dependent production of DXP is ultimately
coupled to the oxidation of NADPH to NADP$^+$ via the DXR enzyme. The solution for the DXS-DXR coupled contained 100 mM HEPES pH 8.0, 100 mM NaCl, 1 mg/mL BSA, 1 mM TDP, 1.5 mM MnCl$_2$, 2 mM β-Me, 0.15 mM NADPH, 0.2 mg/mL DXR, and varying concentrations of pyruvate or G3P$^7$. Steady-state kinetic experiments were performed by varying pyruvate or G3P at a fixed saturating concentration of the co-substrate. A DXS-DXR reaction solution was incubated at 37°C for 5 min, the reaction was initiated by addition of 358 nM DXS, and the progress of the reaction monitored spectrophotometrically at 340 nm for the oxidation of NADPH. The DXS and DXR employed in this assay were over-expressed and purified based on the methods presented in the supporting information. Each sample was stored at -80°C until used for the assay. The steady state initial velocity for DXS measured at various concentrations of pyruvate and G3P were fit to equation 1 (see SI for plots) using nonlinear regression analysis in Sigma-Plot 12.0.

$$v = \frac{V_{max}[S]}{K_M + [S]} \tag{2.1}$$

2.5 Results and Discussion

A central aim of this investigation is to determine and characterize the mechanism of TDP activation in DXS (Figure 2.2). There are two hypothesized mechanisms acting by a different GB: WMM (Wat9709) and DHM (His434). Though most TK enzymes are thought to rely on a histidine residue as the GB, key structural differences and mutagenesis results suggest DXS might diverge from the majority of TK enzymes$^{20,54,60,129,141}$. The reactant state QM/MM minimized structures (Figure 2.4a, 2.4b) provides some initial insight into this process. Coordination of the water oxygen to the $H_n$ (2.0Å,Figure 2.4b) suggests water could act as the GB. Further, the distance between $H_o$ and $N_e$ (1.8Å, Figure 2.4b) suggests that this could be the final destination of this proton. Alternatively, in the absence of Wat9709, His434 directly interacts with TDP albeit more distantly (4.2Å, Figure 2.4a)$^{197}$. 
Figure 2.4: Representations of the RS for DHM (a) and WMM (b). The dashed black lines illustrate the proton transfer reaction.

To determine each mechanism’s feasibility, the RPATh+RESDi technique was employed and respective minimum energy pathways were computed. A plot of $\Delta E$ with respect to $\delta$ (Figure 2.5) values illustrates the energetic favorability of the WMM over the DHM. A $\delta$ value of 0.3Å corresponds to the TS of both mechanisms. The difference between barriers can partially be explained by a conformational change that occurs during the DHM (Figure 2.6). This involves the movement of His434 into a conformation more favorable for deprotonation of the $4'$-amino moiety. His434’s movement induces a strain in the protein backbone and perturbs the configuration of the local environment. This change in configuration accounts for a portion of the energetic differences between the WMM and DHM but does not provide a complete explanation. Further, a $\Delta E_{WMM}^{\dagger}$ value of 22.7 kcal·mol$^{-1}$ is considerably higher than one might expect for an enzyme catalyzed proton transfer and cannot be explained by a simple conformational change.$^{142}$

The reaction pathway calculations applied a restraint to the proton transfer involved in the DHM or WMM. No other restraints were applied to the system. Upon examination of structural changes during the reaction, a second proton was observed to spontaneously transfer from E373 to the N1 atom of TDP’s AP ring in both mechanisms (Figure 2.7). Since E373 was included in the QM region, the proton transfer occurred in response to electronic changes encountered during each mechanism. The combination of the re-
Figure 2.5: Minimum energy profiles computed for the WMM and DHM. The different x-axes are used because of differences in the reaction coordinate ranges for WMM vs DHM; both are associated with the same y-axis. The ∆E\textsuperscript{‡} are 22.7 kcal·mol\textsuperscript{-1} and 33.7 kcal·mol\textsuperscript{-1} for the WMM (gray circles) and DHM (black squares), respectively.

strained reaction path proton transfer and unrestrained E373 to N1 atom proton transfer represents the tautomerization of the AP to IP state (Figure 2.3). The formation of

Figure 2.6: Representative conformational changes between the RS (yellow) and TS (green) of the DHM.

the ylide state is dependent upon first forming the IP state. There is some debate in the literature over the exact details of the IP state formation (\textit{vide supra})\textsuperscript{8,9,133,141}. Most studies propose an equilibrium between the AP, APH\textsuperscript{+}, and IP TDP states (Figure 2.3)
particularly for apo enzymes. As highlighted in the introduction, the pK\(_a\)s of TDP’s N1 atom and E373 residue (see SI and Introduction) are approximated to be close to one another using experimental and empirically based computational techniques. The combination of the pK\(_a\)s and observed responses from QM/MM calculations suggests a concerted mechanism as previously thought. Additional studies are underway to more fully address this unresolved question.

Figure 2.7: Illustration of the proton transfer from E373 to TDP’s AP ring during the tautomerization reaction. (a) and (b) represent the reactant and product states, respectively. While this figure only depicts the structures of the WMM, a similar response was observed during the DHM.

The CPA method, which approximates electrostatic contributions of a single active site residue, was used to determine the stabilizing/destabilizing effects of active site residues as a function of both states (i.e., RS vs TS) and mechanisms (i.e., WMM vs DHM). Negative \(\Delta\Delta E\) values indicate that a particular residue is more stabilizing towards the TS; whereas positive \(\Delta\Delta E\) values show stabilization of the RS. From the 82
Table 2.1: ΔΔE values for four residues of interest in the WMM and DHM. Negative ΔΔE values indicate preferential stabilization of the TS; while positive ΔΔE show stabilization of the RS preferentially. All values are in kcal·mol\(^{-1}\).

<table>
<thead>
<tr>
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<th>ΔΔE(_{\text{DHM}})</th>
<th>ΔΔE(_{\text{WMM}})</th>
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<tbody>
<tr>
<td>K101</td>
<td>2.0</td>
<td>7.5</td>
</tr>
<tr>
<td>H51</td>
<td>-10.1</td>
<td>-1.2</td>
</tr>
<tr>
<td>K289</td>
<td>-12.3</td>
<td>-2.5</td>
</tr>
<tr>
<td>D430</td>
<td>-21.3</td>
<td>-10.8</td>
</tr>
</tbody>
</table>

active site residues examined, there were 4 that showed substantive differences (Table 2.1). Residues found stabilizing the TS were D430, K289, and H51, and Wat10307. K101 were found to preferentially stabilize the RS. K101, H51 and D430 were found in a

Figure 2.8: Active site conformation of the residues discussed in the CPA results. Images show both the RS (yellow) and TS (green). (a) illustrates the DHM while (b) shows the WMM.
catalytic triad-like configuration in the active site (Figure 2.8). It is unlikely that they play a direct role in this reaction due to their distance from the site of activity (7.2Å). A cluster of water molecules were found to span the distance between the reaction site and triad; which suggests an electrostatic role. K289 coordinates to the negatively charged phosphate tail of TDP (Figure 2.8) and is highly conserved in *D. radiodurans*, as well as other TDP dependent enzymes. In fact, most TDP dependent enzymes are found to require a divalent metal ion and positive residues near the phosphate tail to anchor the cofactor.

The magnitude of $\Delta \Delta E_{\text{DHM}}$ values were consistently larger than the magnitudes of $\Delta \Delta E_{\text{WMM}}$ values. This behavior is attributed to the structural change that the DHM TS must adopt in order to position His434 for deprotonation of TDP’s 4’-amino group. The increased TS stabilization for this mechanism suggests the enzyme is tuned to accommodate alternative activation routes although they may not be the most favorable. For example, active site mutations are a common way that bacteria and other lower life forms (i.e., those that rely on MEP pathway) can adapt to changes in chemical environments. By tuning the DXS active site to stabilize TDP activation via varying general bases, evolutionary fitness is maximized.

To better characterize long range electrostatic effects, QM/MM dipole moments for the RS and TS for each mechanism were computed and visualized (Figure 2.9). The RS dipole moments of both the WMM and DHM were essentially the same. Further, WMM dipoles, both RS and TS, are indistinguishable (Figure 2.9b) whereas the DHM TS dipole moment is significantly perturbed (Figure 2.9a). Again, this effect is attributed to the conformation change His434 undergoes during the DHM and appears to be the underlying source of DXS’s ability to stabilize non-water mediated TDP activation.

Herein, we also report experimental kinetics studies of pyruvate and G3P binding and reaction in DXS and several DXS mutants (Table 2.2). For H434A, there exists negligible increase in catalytic rate for pyruvate as well as G3P in comparison to wild-type, respectively. $K_M$ values also slightly increased by 6.1 and 4.6 folds, respectively. G3P’s negatively charged phosphate tail is thought to bind in a positively charged region of the active site; which contains the polar H434 residue. Additionally, the negatively charged pyruvate is thought to interact with the same positive region but, not as strongly.
Therefore, the mutagenesis results suggest that the electrostatic effects that accompany
the H434A mutation have a clear destabilizing effect on substate binding while enhancing
turnover. This behavior is contrary to what would be expected if H434 is required for
initial TDP activation. Thus, the H434A mutant supports the conclusion favoring a
WMM for TDP activation.

Another interesting correlation between CPA and mutagenesis results is related to the
D430A mutant. As previously discussed, D430 is found in a electrostatic triad of residues
that includes K101 and H51 (Figure 2.8). While the $k_{cat}$ for D430A mutant remains rela-
tively unchanged, the $K_M$ for pyruvate and G3P increases 1.9 and 2.4 times, respectively.
This behavior indicates a role in substrate binding rather than catalysis, similar to H434. The corresponding residue in yeast TK (D477) has been studied previously\textsuperscript{135}. D477 was shown to have a rather large effect on activity and substrate binding. In comparison, DXS shows only a 50% loss of activity that is caused by decreased substrate affinity. This speaks to the difference between DXS and other TK enzymes and highlights the need to study this unique subclass of enzyme.

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<th>Pyruvate</th>
<th>G3P</th>
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<tbody>
<tr>
<td></td>
<td>K\textsubscript{M} (mM)</td>
<td>k\textsubscript{cat}/K\textsubscript{M} (s\textsuperscript{-1}M\textsuperscript{-1})</td>
</tr>
<tr>
<td>Wild-type</td>
<td>0.28 ± 0.03</td>
<td>2.6 × 10\textsuperscript{4}</td>
</tr>
<tr>
<td>H82A</td>
<td>0.23 ± 0.02</td>
<td>1.7 × 10\textsuperscript{3}</td>
</tr>
<tr>
<td>H304A</td>
<td>1.7 ± 0.5</td>
<td>5.8 × 10\textsuperscript{2}</td>
</tr>
<tr>
<td>D430A</td>
<td>0.52 ± 0.5</td>
<td>1.4 × 10\textsuperscript{4}</td>
</tr>
<tr>
<td>H434A</td>
<td>1.7 ± 0.1</td>
<td>5.9 × 10\textsuperscript{3}</td>
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<tr>
<th></th>
<th>K\textsubscript{M} (mM)</th>
<th>k\textsubscript{cat}/K\textsubscript{M} (s\textsuperscript{-1}M\textsuperscript{-1})</th>
<th>k\textsubscript{cat} (s\textsuperscript{-1})</th>
<th>%WT</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wild-type</td>
<td>0.05 ± 0.01</td>
<td>1.5 × 10\textsuperscript{5}</td>
<td>7.9 ± 0.4</td>
<td>100</td>
</tr>
<tr>
<td>H82A</td>
<td>0.03 ± 0.01</td>
<td>1.3 × 10\textsuperscript{4}</td>
<td>0.37 ± 0.02</td>
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</tr>
<tr>
<td>H304A</td>
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<td>1.1 × 10\textsuperscript{4}</td>
<td>0.90 ± 0.1</td>
<td>11.4</td>
</tr>
<tr>
<td>D430A</td>
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<td>6.6 × 10\textsuperscript{4}</td>
<td>7.7 ± 0.2</td>
<td>97.5</td>
</tr>
<tr>
<td>H434A</td>
<td>0.23 ± 0.01</td>
<td>4.2 × 10\textsuperscript{4}</td>
<td>9.6 ± 0.3</td>
<td>121.5</td>
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Table 2.2: DXS steady-state kinetics data (wild-type and mutants) for both pyruvate and G3P. %WT was determined by comparing the mutant k\textsubscript{cat} to the wild-type k\textsubscript{cat}.

Two histidine residues are in close proximity to each other (3.7Å between N\textsubscript{e} atoms for H82 and H304) and the center of activity (5.1Å and 5.7Å from the thiazolium C2 atom respectively for H82 and H304) of DXS. Table 2.2 shows that the H82A and H304A mutants produce catalytically defective enzymes resulting in only 2-12% k\textsubscript{cat} and k\textsubscript{cat}/K\textsubscript{M} values when compared to wild-type. The loss of activity can be explained by their proximity to the thiazolium C2 atom. These residues can assist in stabilizing the α-carbanion\textbackslash enamine intermediate following pyruvate decarboxylation (Figure 2.2, step 5). While activity in these mutants is significantly retarded, detectable levels of activity are retained. This retention might be explained by the proximity of these two residues to one another. Upon the loss of one histidine, it is possible for the other His residue to recover partial functionality. There is one noticeable difference in the results of these two mutants. The H304A K\textsubscript{M} for pyruvate has increased compared to the wild type; while K\textsubscript{M} value for H82A
remain similar to the wild-type value. This indicates that while both residues are clearly catalytically important, H304A protrudes into the pyruvate binding site and, therefore, plays a role in binding; which can not be replaced by H82. Thus, accounting for observed differences in mutant $K_M$ values for pyruvate.

Figure 2.10: Analysis for 18 ns of the unrestrained simulation of the 2O1X DXS structure utilized in this investigation. (a) the distances over time for $N_\epsilon$ of H434 to N4’ of TDP’s amino group. (b) shows the fluctuations for backbone (black), and side chains (grey) for residues H51, K101, H124, K289, E373, D430, and H434. These residues represent the QM region and key CPA residues previously discussed. (c) snapshot from the 18 ns trajectory with H434 in proximity to the 4’-amino group. (d) is representative of H434 in the second conformation.

With these mutagenesis results, it became apparent that a longer simulation was required to examine active site conformational dynamics. Thus, the 2O1X structure was simulated for an additional 20 ns with the first 2 ns discarded (details found in supplementary information). The trajectories were compared to the QM/MM minimized RS. The distance between the 4’-amino group and the $N_\epsilon$ atom of H434 revealed two major conformations (Figure 2.10a). The first conformation lasts for $\sim5.0$ ns and has H434 3.5Å from the AP ring on average. The second conformation has H434 7.1Å from the AP ring on average and remains throughout the simulation. The fact that the 2nd conformation is stable for the majority of the simulation and places the histidine beyond the range of
direct deprotonation of its 4′-amino group provides further support for a water mediated mechanism. Additionally, the backbone and side chain fluctuations were calculated for significant CPA residues (e.g., E373 and H434, Figure 2.10b). The conformational change of H434 to a position proximal to K101 and D430 accounts for the larger side chain fluctuations of K101 and D430 (Figure 2.10). The introduction of H434’s imidazole would force K101 and D430 to move in order to accommodate the bulky polar side chain. The combination of the motion of these residues with H434 being the final resting place of the proton abstracted from TDP’s amino group suggests a possible regulatory role for H434. H434 could act as a shuttle involved in regenerating the TDP-ylide for further reactions by displacing the abstracted proton onto D430. This perfectly aligns with experimental results showing that the removal of this residue (H434A) slightly increases $k_{cat}$, allowing any proton transfer from TDP’s amino group to D430 to occur more rapidly via a water mediated process (picosecond time scale) rather than the H434 side chain motion that likely occurs on the nanosecond time scale.

<table>
<thead>
<tr>
<th>NICS</th>
<th>DHM-RS</th>
<th>DHM-TS</th>
<th>∆NICS</th>
</tr>
</thead>
<tbody>
<tr>
<td>(1)</td>
<td>-7.1</td>
<td>-5.7</td>
<td>1.4</td>
</tr>
<tr>
<td>(0)</td>
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<tr>
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<td>-9.3</td>
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**Average**  

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<th>WMM-TS</th>
<th>∆NICS</th>
</tr>
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</tr>
<tr>
<td>(-1)</td>
<td>-8.8</td>
<td>-6.3</td>
<td>2.5</td>
</tr>
</tbody>
</table>

**Average**  

Table 2.3: Calculated NICS values for the WMM and DHM RS and TS. The NICS(0) values are taken from the center of the pyrimidine ring. The NICS(1) and NICS(-1) values are points away and towards a proximal phenylalanine (F398), respectively. A comparison set of benzene (-9.8) and cyclobutadiene (27.6) were computed to show reference aromatic and antiaromatic values, respectively. The average ∆NICS values represent a 29.4% and 35.6% decrease in aromaticity for DHM and WMM, respectively.

TDP reactivity is clearly dependent on the surrounding environment, e.g., rate of reaction increases a billion-fold when bound to an enzyme\textsuperscript{84}. Several attempts to determine the underlying energetics have attributed this behavior to the strained ‘V’ shape
TDP adopts upon binding\textsuperscript{25,26,82}. Given the pK\textsubscript{a} changes this conformation induces, it is surprising that the barrier to activation (i.e., proton transfer) is significantly higher than expected; $\Delta E^\dagger=22.7$ kcal mol\textsuperscript{-1} vs. 5-10 kcal mol\textsuperscript{-1} for typical proton transfers\textsuperscript{142}. One possible cause of this is the loss of aromaticity that occurs during ylide formation. Additionally, overestimation of the barrier may be due to the inability to carry out free energy simulations; nevertheless, the energy barrier difference is a more meaningful quantity when seeking to differentiate two possible mechanisms.

To examine the former, i.e., aromaticity effects, both NICS and ASE were computed. NICS calculations estimate the aromaticity of a molecule; negative NICS values indicating aromaticity and positive NICS values antiaromaticity. Table 2.3 reports NICS results for RS and TS of TDP activation via DHM and WMM. An average of the $\Delta$NICS values was used to quantify the relative change in aromatic character. Results indicate the AP ring is aromatic in both the RS and TS with values close to those published for similar pyrimidine analogs\textsuperscript{1}. However, the TS consistently shows lower aromatic character than the RS; which supports our hypothesis of aromaticity regulating ylide formation.

Calculating the ASE for TDP's AP state should provide additional information about the importance of aromaticity in ylide formation. A homodesmotic reaction (Figure 2.11) provides a reference for determining ASE. Thiamin serves as a model compound for this purpose and represents the key components (e.g., 4'-amino and thiazolium moieties) of TDP. Systems with positive values of ASE are considered to be aromatic, whereas those with negative values are antiaromatic. ASE values are determined as the difference in energies between both halves of the reference reaction (Figure 2.11). Thiamin has an ASE

![Figure 2.11: Homodesmotic reaction used in evaluating the aromatic stabilization energy for a model TDP.](image-url)

\textbf{Figure 2.11:} Homodesmotic reaction used in evaluating the aromatic stabilization energy for a model TDP.
of 37.6 kcal·mol$^{-1}$; which is again in close agreement with previously published results of similar pyrimidine derivatives$^1$.

Combining ASE values with the average decrease in aromaticity (i.e., $\Delta NICS$, Table 2.3), we approximate the stabilization lost at the TS of each mechanism. Aromaticity losses of 13.4 kcal·mol$^{-1}$ and 11.1 kcal·mol$^{-1}$ for WMM and DHM were computed, respectively. Interestingly, we again observe the DHM TS being less destabilized when compared to the WMM TS. This provides further evidence that DXS is well adapted to stabilizing alternative mechanisms of TDP activation. Finally, when the total barrier heights are considered it becomes clear that the loss of aromaticity plays a major role in TDP activation and the initial step of isoprenoid biosynthesis.

2.6 Conclusion

The $\Delta \Delta E^\ddagger$ of 10.0 kcal·mol$^{-1}$ difference between the WMM and DHM mechanisms indicates the WMM is the energetically favorable route for ylide formation in DXS. The RPATh+RESDi results seem to suggest the mechanism of proton transfer acts in a concerted fashion proceeding via the tautomeric route between the AP and IP state. Further investigation is ongoing to confirm the relative energetics of a step-wise versus concerted mechanism.

CPA results were indicative of H434 playing a role in long range electrostatic stabilization; which is more clearly illustrated upon examination of the RS and TS active site dipole moments. Mutagenesis studies performed reveal H434 to play a role in substrate binding but not likely a direct role in catalysis. The H434A mutant results reinforce the CPA results. Additionally, a D430A mutant revealed a lower catalytic significance for DXS in comparison to the corresponding yeast TK mutant$^{135}$; again illustrating mechanistic differences. Furthermore, H82A and H304A DXS mutants showed significant decreases in activity (2-12% of wild-type). Given their proximity and retention of measurable activity, it is likely these residues function as back-ups to each other. This comports nicely with computational results that indicate DXS is well suited to functioning via alternative mechanisms (i.e., different general bases), something that would offer a significant evolutionary advantage.
$\Delta E^{\ddagger}_{WMM}$ is significantly higher than what might be expected for a relatively simple proton transfer\textsuperscript{142}. CPA results could not account for such behavior. However, upon examination of the 4$'$-amino moiety, it was evident from structural changes that aromaticity may be changing. The results of NICS and ASE calculations showed that indeed the AP ring was losing aromaticity. If the aromatic contribution is taken into consideration, the new $\Delta E^{\ddagger}_{WMM}$ would be closer to 8.5 kcal-mol$^{-1}$; which is in the range of similar reactions. This clearly shows that loss of aromaticity plays a key role in controlling activation of TDP in DXS. Further, restoration of this aromaticity upon intramolecular proton transfer from the C2 of the thiazolium ring to the 4$'$-amino group should ultimately drive the final ylide formation.

2.7 Supporting Information (SI)

The following can be found in Appendix A: PDB files for the transition states of the WMM and DHM; link atom details for QM/MM reaction path calculations; CPA results; topology and parameter files for TDP; methods for the over-expression and purification of DXS and DXR; steady-state initial velocity plots with varying concentrations of G3P and pyruvate; details about 20ns simulations; results of PROPKA3.1 calculations on DXS Chain A.

2.8 Acknowledgments

H.L.W. would like to acknowledge NIH (1K22HL088341-01A1, 4K22311045-02), NSF (CHE-1464946), and the University of South Florida (start-up) for funding. Additionally, D.J.M. would like to acknowledge funding from two NIH grants (RO3-DA034323 and R15-GM107864). Computations were performed at the USF Research Computing Center (NSF Grant No. CHE-1531590). This research was supported in part by a seed grant from the Florida Center of Excellence for Biomolecular Identification and Targeted Therapeutics (FCoE-BITT) to D.J.M. and a Graduate Multidisciplinary Scholar (GMS) award from FCoE-BITT to S.H.
Chapter 3

Computational Examination of the Magnesium Ion Binding Modes of
1-Deoxy-D-xylulose 5-Phosphate Reductoisomerase

3.1 Introduction

There exists a vast and varied class of natural products derived from two five-carbon iso-
prene precursors, isopentenyl diphosphate (IDP) and dimethylallyl diphosphate (DMADP),
and serve several essential roles for all living organisms\textsuperscript{144}. These are generally known as
Isoprenoids. The broad variety of uniques molecules comprising this family are derived
via a combination of elongations, rearrangements, cyclizations, and oxidations utilizing
IDP and DMADP in various combinations\textsuperscript{38}. A few of the important biological roles
filled by isoprenoids are prenyl lipids in archaebacteria\textsuperscript{46}, sterols in eubacteria and eu-
karyotes\textsuperscript{128}, light-harvesting pigments such as carotenoids, electron transport carrier such
as ubiquinone and menaquinone, and several growth and development regulators (Figure
3.1)\textsuperscript{161}. Additionally, there are several known herbicides or herbivore repellents identified
to be isoprenoids\textsuperscript{48}.

The biosynthesis of the IDP and DMADP building blocks were originally thought
to derive from a single enzyme pathway (Figure 3.1\textsuperscript{132}. This pathway is known as the
mevalonate dependent (MVA) pathway; which was named after the key committed inter-
mediate formed from the condensation and reduction of 3 acetyl-CoA molecules producing
mevalonic acid (or mevalonate in ionic state). Continuing discrepancies in the results of
isotope labeling studies\textsuperscript{30,31,57,140,205} led several researchers to postulate the existence of
a second yet unidentified pathway. Efforts by researchers such as Rohmer, Arigoni, Lich-
tenthaler, and Seto, *etc.* eventually discovered a new pathway completely distinct from the MVA pathway\textsuperscript{32,91,111}. Initially, the pathway had names reflecting the distinction between MVA and the novel, mevalonate-independent or non-mevalonate (NMA) pathway. Seven enzymes catalyzing 8 reactions comprise the NMA pathway\textsuperscript{132}. Rohmer and co-workers\textsuperscript{159} established the formation of 1-deoxy-D-xylulose 5-phosphate (DXP) via the decarboxyl condensation of pyruvate and glyceraldehyde-3-phosphate (G3P). DXP is additionally utilized as an intermediate for the biosynthesis of vitamins B1 and B6 as well as isoprenoid biosynthesis\textsuperscript{114,160}. Therefore, DXP is required and considered the first step of the pathway but not the committing step in the NMA pathway. The succeeding reaction catalyzed by DXP reductoisomerase (DXR) bares the distinction of being
the committed step in the NMA pathway. The product of this reaction, 2-C-methyl-D-erythritol 4-phosphate (MEP), lends its name to the pathway, as the NMA pathway is frequently referred to as the MEP pathway\textsuperscript{100}.

DXP reductoisomerase catalyzes a carbon-skeleton rearrangement of DXP and subsequently reduced. DXR activity requires a divalent metal cation cofactor and NADPH co-substrate. Out of the divalent metal ions attempted, DXR is activated by only Mn\textsuperscript{2+}, Co\textsuperscript{2+} and Mg\textsuperscript{2+}, in decreasing order respectively\textsuperscript{3,94,102,118,179,202}. Additionally, NADH was tested in place of NADPH as the co-substrate with DXRs derived from \textit{E. coli}, \textit{M. tuberculosis} and \textit{S. leopoliensis}. The results were a decrease in activity in some cases as much as a 100-fold decrease in activity. Decreased affinity is responsible for the lost activity primarily due to the loss of the 2\textsuperscript{′}-phosphate of NADPH. Therefore, the phosphate is a binding determinant and not likely directly involved in catalysis since \( k_{\text{cat}} \) was unaffected\textsuperscript{3,179}.

Results of isotopic labeling studies demonstrated the required isomerization proceeding via a C3/C2 bond transition. The isomerization results in an aldehyde intermediate 2-C-methyl-D-erythrose 4-phosphate (MEsP), which is subsequently reduced on the \textit{re} face of MEsP by the C4 pro-S hydride of NADPH\textsuperscript{4,5}. The proposal of this intermediate was originally based on analogy to ketol-acid reductisomerase (KARI), which catalyzes a similar reaction during the biosynthesis of branched-chain amino acids. As with the KARI reaction, the MEsP aldehyde intermediate has never been directly detected\textsuperscript{50,100,179}. Several attempts have been made to isolate the aldehyde intermediate with no success. These results suggest the intermediate might be more transient than originally thought or very tightly bound prior to NADPH reduction\textsuperscript{78,179}; which has been similarly proposed for KARI\textsuperscript{50}. Rohmer and co-workers produced the first compelling evidence supporting the aldehyde intermediate theory by introducing exogenously synthesized MEsP and demonstrating kinetic competency. When incubated with DXR in the presence of NADPH and Mg\textsuperscript{2+} or Mn\textsuperscript{2+}, a factor of 4 and 1.6, respectively, increase in conversion to MEP was observed. While the oxidized coenzyme was present, a 7\% conversion of MEsP to DXP
was detectable\textsuperscript{37}. Additionally, the $K_m$ for MEsP was found to be greater than DXP by a factor of 4 and 1.6 in the presence of Mg\textsuperscript{2+} or Mn\textsuperscript{2+}, respectively. The argument has been made based on these values against the tight-binding of MEsP. This is flawed though since $K_d$ and $K_m$ are only equal when substrate dissociation is rapid\textsuperscript{78,137}.

Despite the overall similarities of DXR- and KARI-catalyzed reactions, amino acid differences suggest different mechanism of action\textsuperscript{37,49,95}. Three mechanisms were proposed to explain the carbon-skeleton rearrangement: 1) an $\alpha$-ketol rearrangement, 2) a retro-aldolization/aldolization, and 3) a sequential 1,2-hydrdride and 1,2-methyl shift\textsuperscript{64}. A dismissal of the third mechanism was accomplished based on $^{13}$C-glucose and $^{13}$-DXP incorporation studies\textsuperscript{3,72}. Therefore, further investigations looked to distinguish between the remaining $\alpha$-ketol rearrangement or retro-aldol/aldol mechanism (Figure 3.2).

The retro-aldol/aldol mechanism should form 2 putative intermediates of glycoaldehyde phosphate and the enolate of hydrxyacetone. If these intermediate could be detected during or following the reaction, it would provide strong evidence in support of the retro-aldol/aldol mechanism. Several attempts were made with no success\textsuperscript{58,78,104}. Though the lack of detection is consistent with both mechanisms as the results can be explained as the intermediates fragments are tightly confined to the active site. Additionally, these putative fragments could be so transient, they never truly form. Subsequent, experiments have tended to favor the retro-aldol/aldol mechanism, such as the modification or removal of the C4 hydroxyl group. The $\alpha$-ketol rearrangement doesn’t require the C4 hydroxyl group and therefore any turnover would support. Though turnover was not observed for 1,4-dideoxy-D-xylulose 5-phosphate, the $K_i$ values similar to the $K_m$ indicates a dependence on the C4 moiety for turnover but not binding. The C4 epimer and fluorinated version of DXP produces similar results\textsuperscript{143,193}. Due to the relatively good binding of these modified ligands, the retro-aldol/aldol mechanism is favored.

The analogues studies have provided some significant evidence in support of the retro-aldol/aldol mechanism over the $\alpha$-ketol rearrangement. Kinetic isotope effects (KIEs) provide a means of further probing the mechanism. In order to differentiate between
Figure 3.2: The above illustration compares the steps for the α-ketol rearrangement and retro-aldol/aldol mechanisms in a side-by-side view. Each concludes in the aldehyde intermediate 2-C-methyl-D-erythrose 4-phosphate (MEsP), which is subsequently reduced by NADPH to form the 2-C-methyl-D-erythritol 4-phosphate (MEP) product.

The two mechanisms, α-secondary KIEs were measured for [3-²H]- and [4-²H]-DXP. The α-ketol rearrangement predicts a shift from sp³ to sp² at the C3 position while the C4 position remained sp³, which translates into KIEs>1 and unit KIE values, respectively.
In contrast, the retro-aldol cleavage both C3 and C4 undergo changes from sp$^3$ to sp$^2$ with normal KIE values (KIE>1)$^{131}$. The results of 1.04 for [3-2H] and 1.11 for [4-2H]-DXP supports the retro-aldol/aldol mechanism. When compared to muscle aldolase, which has a similar mechanism, the lower KIEs are thought to reflect the partially rate-limiting rearrangement or an early transition state$^{131}$. Finally, a 2D [$^{13}$C,$^1$H]-HSQC NMR based technique was used to analyze $^{13}$KIEs. The method measures the reactive competition between light and heavy C substrates in the same mixture with the enzyme. The ratio of $^{13}$C/$^{12}$C represents the KIE. The ratios were measured for 2-, 3-, and 4-$^{13}$C with results of 1.0031, 1.0303 and 1.0148, respectively$^{120}$. The sigmotropic rearrangement would result in large changes at all locales while retro-aldol predicts larger changes at the C3 and C4 position with little effect on the 2C position. The results of these KIE experiments supports the retro-aldol/aldol mechanism as the most likely mechanism. The only major issue left to challenge the retro-aldol/aldol mechanism is the failure to detect the putative hydroxyacetone and glycoaldehyde intermediates. Currently, the best explanation revolves around the tight binding of these intermediates and/or the molecules exist in such a high energy state, they aren’t around long enough to be a true intermediate$^{132}$.

Figure 3.3: Illustration of the C2-C3 and C3-C4 binding modes in the reactant state. These structures were utilized for the purposes the replica path calculations as the starting point.
The retro-aldol mechanism was originally proposed with the metal ion coordinated between the C2-C3 hydroxyl groups. Results of incubating DXR with Mg\textsuperscript{2+}, NADPH and DXP in the presence of \textsuperscript{18}O-labeled water to explore the incorporation of the isotope into MEP\textsuperscript{109} dispute this proposal. Retro-aldol/aldol mechanism produces carbonyls, if transiently, at each position during the reaction, thus allowing for solvent exchange at the C2, C3, and C4 positions of DXP. Since the only hydroxyl affected was the C2 of DXP, there had to be a protective effect at the C3 and C4 positions. Coordination of the divalent ion would act as protection from solvent exchange, therefore the results suggest a C3-C4 binding mode. This binding mode helps to explain the tight binding of both fragments since the retro-aldol cleavage occurs along the C3-C4 bond\textsuperscript{74,118,172,178}. During the bond breaking and subsequent C2-C4 bond forming steps, the Mg\textsuperscript{2+} would remain coordinated to both fragments inhibiting release. It is still possible to interpret the results of these experiments in support of the C2-C3 binding mode. The Lewis acid characteristics of the metals would increase the electrophilicity of the C2 carbonyl thus promoting hydration. Furthermore, the transiency of the intermediates may explain the lack of solvent exchange. Exchange of the C1 or C3 oxygen atoms requires the rate of on-enzyme hydration to rival rates of hydride transfer and aldolization\textsuperscript{132}.

Examining the energetics of the metal binding modality will be the focus of this work. Mac Sweeney et. al. published a crystal structure of \textit{E. coli} DXR (PDB:1Q0Q) with DXP and NADPH bound in the active site\textsuperscript{118}. The experimental results published to this point provide strong support for the retro-aldol/aldol mechanism being the most likely reaction mechanism, so it was decided to focus on this pathway for our calculations. In particular, we focused on the retro-aldol calculation, which is thought to be the true limiting step of this reaction. The putative intermediates are even thought to not be proper intermediates but possibly transition states. QM/MM techniques were utilized to compute the free energy surface of the retro-aldol reaction with the metal ion in the C2-C3 or C3-C4 position.
3.2 Computational Methods

The crystal structure published by Mac Sweeney et. al. (PDB:1Q0Q) was utilized for all calculations in this paper \(^{118}\). Although, DXR is generally found to be in a homodimer in solution, there is no evidence currently supporting catalytic interdependence of active sites. Thus, allowing up to focus on a single monomer. The structured was parsed utilizing www.charmming.org \(^{127}\). Parameters for DXP were built based on similar structures already found in the CHARMM General Force Field (CGenFF) \(^{184}\). The necessary bonds were added based on the most similar structures and the charges were corrected via quantum mechanical calculation. Final validation was performed utilizing the crystal structure as the comparison.

The protein was built and E234 was protonated based on values determined by ProPKA3.1 (see Appendix B) \(^{138,175}\). CHARMM22 protein and CGenFF force fields were used throughout these calculations \(^{119}\). A Mg\(^{2+}\) ion was built separately and added to the composed enzyme. The ion was brought into C2-C3 and C3-C4 orientation via use of the harmonic distance restraint (RESDi) while fixing the rest of the system followed by an unrestrained minimization. The system was solvated in a rhombododechedron crystal structure and neutralized with KCl salt to a final concentration of 0.15M. The system was heated from 110K to 310K over 100ps and equilibrated for 200ps at constant pressure (1atm) and temperature (310K). The total system size was subsequently reduced to cut down on computational costs in the following QM/MM calculations by removing all waters/ions beyond 12Å from the protein surface. The reduced structure was treated to a QM/MM minimization without cut-offs to a tolerance of 0.002 kcal·mol\(^{-1}·Å^{-1}\). All QM/MM calculations employed the Q-Chem4.0\(\text{\textbackslash }\)CHARMM\(^{27,170,194}\) interface at the B3LYP/6-31G* level of theory \(^{13,68,105}\). Additionally, the single link atom scheme was used to account for truncation of the QM region and employed group electrostatic exclusions to prevent over polarization of the QM region (23 atoms) \(^{167}\). The QM region was defined as the D231, MG, and DXP only. The NADPH molecule was excluded because it is not thought to play a role in the skeletal rearrangement.
Reaction path calculations were performed using the Replica Path (RPATh) in combination with RESDi values to define the steps along the reaction coordinate. RPATh allows the user to define a subsection of the structure which will be duplicated into replicas. These replicas are free to react normally as the reaction progresses while the larger system is constrained thus cutting down on the computational costs. The replicas utilized in these calculations were included the QM region and a buffer region of 6.5Å around the QM section. For our purposes, two replicas were utilized. In order to provide a buffer from the constrained system, the replicas were defined as all residues within 6.5Å from the QM region. One replica was incrementally progressed along the reaction. This was performed by defining two reaction coordinates ($\delta_1$, $\delta_2$); which were defined with reference to reaction component being controlled.

$$\delta_1 = Bond - Breaking_{C3-C4}; \delta_2 = Bond - Breaking_{O4-H9} - Bond - Forming_{H9-OE2}$$ (3.1)

As the retro-aldolization is composed of two parts, $\delta_1$, corresponding to the breaking of DXP’s C3-C4 bond, could be easily be defined while $\delta_2$, corresponding to the deprotonation of the C4 hydroxyl by residue D231, was defined as a linear combination of distances. A two-dimensional energy surface was produced with respect to these reaction coordinates. While the progression along $\delta_2$ was easily defined as beginning at -2.0 (reactant state) and ending at 2.0 (intermediates) as an assumption like previous work, the path of $\delta_1$ was more difficult. Since $\delta_1$ refers to a single bond breaking, the C3-C4 bond of DXP was elongated by 2.0Å. $\delta_2$ was progressed in 0.2Å increments and $\delta_1$ was allowed 0.1Å increments for a 21x21 point 2D surface. After the completion of these calculations, normal mode analysis was utilized to identify the reaction steps corresponding to states of interest (“products”). This was performed by QM calculations utilizing Q-Chem as “freq” jobs. The output frequencies were analyzed for unique asymmetric vibrations corresponding the changes desired.
3.3 Results and Discussion

The first response to the complete 2D-energy surface indicates our initial ranges for the reaction coordinates may have been too broad. Both the C2-C3 and C3-C4 show a range of values produce very strained structural states. A few of these values were repeated to verify with similar results, therefore the rest of the work focused on results prior to $\delta_2=0.8-2.0$ for C2-C3 and $\delta_2=1.2-2.0$ for C3-C4 calculations. The structures present structures representing over extended CO bonds and massively contorted structures.

Before continuing discussions of the energy results, it is important to discuss the identification of the “products”; which correspond to the putative intermediates between the retro-aldol and aldol steps of the DXR reaction. Systems with unique normal modes were found at $\delta_1;\delta_2=3.40;0.60$ for C2-C3 coordinated state and $\delta_1;\delta_2=3.40-3.60;1.00$ for the C3-C4 metal coordination. This results supports the proposal to exclude the results mentioned previously.

Figure 3.4: Two-dimensional energy surfaces with outlier values removed. With the outliers for the C2-C3 (image (a)) and C3-C4 (image(b)) removed from the surface plot, the details are more easily observed. The valley in the top right of image (b) might indicate a step-wise mechanism.

The results of the reaction path calculations (Figure 3.4) reveal some distinct differences between the binding modes. While the C3-C4 binding mode has distinct peaks and valleys, the C2-C3 surface is ever increasing (Figure 3.4a). This result is not really grounds for dismissal of the binding mode. As the “products” of the retro-aldol reac-
Figure 3.5: One-dimensional representation of the center path across the two-dimensional energy surfaces for the C2-C3 and C3-C4 binding modes.

tion are, in actuality, intermediates or more likely transient transition states during the skeletal rearrangement phase of the DXR reaction. Both binding modes conclude at high energy states (33.0 kcal·mol$^{-1}$ and 30.9 kcal·mol$^{-1}$ for C2-C3 and C3-C4 binding modes, respectively).

For easier comparison, scatter point plots of pathways representing the best path between points were produced (Figure 3.5). The results suggest the binding modes to be rather similar. The barrier energies are 36.0 and 37.6 kcal·mol$^{-1}$ for the C2-C3 and C3-C4 mode, respectively. So overall, the energetics of the C2-C3 compared to C3-C4 isn’t sufficient to address the question binding modes. A structural comparison of the “products” in conjunction with the energetics makes for a different outcome. The retro-aldol reaction needs a deprotonation to activate the breaking of the C3-C4 bond producing the hydroxyacetone enolate and glycoaldehyde phosphate “products”\textsuperscript{132}. The base is proposed to be the D231 residue found in proximity of the DXP hydroxyl groups, which was controlled by the $\delta_2$ reaction coordinate. Figure 3.6 illustrates the “products” states of each binding mode. The C3-C4 binding mode (Figure 3.6b) are a clear representation of the intended hydroxyacetone enolate, glycoaldehyde phosphate and protonated D231 residue “products”. The “product” state for the C2-C3 binding mode (Figure 3.6a) reveals an intermolecular protonation of the phosphate group despite the presence of a
restraint directing proton transfer. There are two possible explanations for this difference configurations. The C3-C4 binding mode held the C4 hydroxyl group in a favorable position for deprotonation by the D231 oxygen. Secondly, the metal, acting as a lewis acid, could have further polarized the O4-H bond, thus promoting deprotonation by the glutamate.

Figure 3.6: The final structures of “products” for the retro-aldol reaction are the hydroxacetone enolate and glycoaldehyde phosphate at the bottom of the two images above. Image (a) above represents the C2-C3 binding mode that results in a intermolecular protonation of the phosphate group and preferential coordination with the C2-C3 oxygens. On the right, image (b) is the C3-C4 binding mode which produced the desired products and protonation states.

Along with the possible structural highlights for the mechanism, figure 3.6 illustrates another key difference in the binding modes. Namely, the position of the metal ion after preparation and RPATh calculations. While the magnesium remains straddling the O2 and O3 atoms of the C2-C3 mode (Figure 3.7a,c), the C3-C4 binding mode actually transitions into an all oxygen coordination (Figure 3.7b). The conformational change occurs spontaneously after the heating and equilibration phases of the build phase. Figure 3.7 reveals the differences in coordination between the pre- and post-equilibration steps for each binding mode. Figure 3.7b shows the Mg$^{2+}$ to rest 2.76 Å from the O2 atom prior to equilibration, which likely represents a local minimum on the energy potential. After the injection of energy from heating and equilibration, the distance from the O2 atom reduces to 2.31 Å. The conformational change observed between these steps occurred
Figure 3.7: Pre-equilibrium and post-equilibrium for each binding mode. Images (a) and (c) represent the C2-C3 binding mode pre-equilibration and post-equilibration, respectively. Images (b) and (d) represent the C3-C4 binding mode pre-equilibration and post-equilibration, respectively. The side-by-side comparison highlights the changes made.

with no restraints on the system. In contrast, the C2-C3 structures show no interesting in reaching out the O4 atom of DXP. The post-equilibrated structure looks like what one might expect. The originally minimized structure shows some leveling out between the coordination bonds.

The putative intermediates, hydroxyacetone enolate and glycoaldehydephosphate, and reaction intermediate, MEsP, have never been observed directly\textsuperscript{78,179}, which is the remaining hope for the \( \alpha \)-ketol rearrangement mechanism. An explanation is the tight binding of the reactants in the active site\textsuperscript{104}. This spontaneous coordination to all DXP oxygens might be further support for the tight binding hypothesis. While the C2-C3 binding mode would only effect the binding of the hydroxyacetone intermediate, the glycoalde-
hyde phosphate might actually be able to leave the active site. Additionally, an all oxygen coordination might aid in the subsequent aldolization by holding all the intermediates together and aiding in stabilizing the C2-C4 bond.

As previously mentioned, the C3-C4 energy surfaces have values with mechanistic implications. Generally, the retro-aldolization requires a deprotonation of the C4 hydroxyl group, and is thought to occur concurrently with the C3-C4 bond breakage (Figure 3.8c). The energetics displayed during these calculations are the first hints of a step-wise retro-aldol reaction beginning with the proton transfer from a highly polarized hydroxyl group and proceeding to the C-C bond breakage. The other valley corresponding to C-C bond breakage occurring first does not contain any structures with unique normal mode frequencies, so this valley is probably an outlier in the data. Figures 3.8a,b are scatter point plots of each proposed step (figure 3.8c, respectively. The $\Delta E^\ddagger$ for the deprotonation of 30 kcal-mol$^{-1}$ is considerably higher than one might expect. Therefore, the $\Delta E^\ddagger$ indicates the deprotonation as the rate-limiting step. The relatively high energy of the deprotonated state produces a reduced $\Delta E^\ddagger$ to the final retro-aldolization “products” produced by C3-C4 bond breakage. These results are far from conclusive but provide a new area of further study.

3.4 Conclusion

The energetics of the two binding modes suggest a preference for the C3-C4 binding mode over the C2-C3 binding mode. It is a slight difference of 2.1 kcal-mol$^{-1}$; which means the energetics aren’t definitive. The combination with the fact the C2-C3 “products” show an intermolecular proton transfer suggests the C2-C3 binding mode to be unfavorable.

In addition to the energetics, configurational differences between the binding modes provide further evidence in support of the C3-C4 binding mode. The spontaneous shift of the C3-C4 mode into a C2-C3-C4 mode enhances the arguments for DXR strongly binding the intermediates; which explains why they haven’t been directly observed. Reaction assistance provided by the expanded binding mode could explain the formation
of the desired “products” unlike the C2-C3 while holding the intermediate in proximity necessary for the aldolization. This aldolization assistance could provide further evidence these intermediates being truly transition states.

Further work should start with analyzing the changes in active site contributions over the reaction. The mapping of the aldol reaction should be performed starting at the “product” state of the retro-aldol reaction for both binding modes to see if there is a change in preference between the stages of the skeletal rearrangement.

### 3.5 Supporting Information (SI)

The following can be found in Appendix B: The ProPKA3.0 results of crystal structure for DXR produced by MacKerrell (PDB:1Q0Q).
Chapter 4

Conclusion and Future Work

The work shown in this document represent the initial steps in gaining understanding of the reactions involved in the NMA pathway for Isoprenoid biosynthesis. The work on 1-deoxy-D-xylulose 5-phosphate (DXP) synthase (DXS) focused on the deprotonation of the N4 atom of the thiamine diphosphate (TDP) cofactor, which occurs in preparation of the formation of the ylide via deprotonation of the C2 atom. This step highlights the significance of QM/MM reaction path calculations. Work similar to this can provide insights into reaction steps of an enzyme or even a portion of a step. Kinetics can provide similar insights but are dependent on the step of interest being the rate-limiting step, which provides no assistance with DXS and other TDP dependent enzymes. The ylide activation step is required for activity but happens at the rate of diffusion so experimental practices are currently ineffective. By utilizing computational techniques, it is possible focus on pieces of a reaction and tweeze out pieces of information of significance. Both experiments in this document deal with half of a reaction commonly referred to as a single reaction because they can’t be measured experimentally.

4.1 1-Deoxy-D-xylulose 5-Phosphate Synthase Summary and Conclusion

The mechanism of DXS consists of 3 major pieces: ylide activation, pyruvate binding, and pyruvate decarboxylation couples with transferral of acetyl group to glyceraldehyde-3-phosphate (G-3-P). The conclusion is the production of DXP; which is utilized in the production of the isoprenoid precursors of isopentenyl diphosphate (IDP) and dimethyl-
lallyl diphosphate (DMADP). The rate-limiting step of the reaction is the pyruvate step. The ylide formation is thought to happen at the rate of diffusion but is required for enzymatic activity since the ylide acts as the reactive center for the enzyme. Prior to activation, a deprotonation happens at the N4 atom of the TDP pyridinium ring producing a tautomeric transition between the 4′-aminopyrimidine (AP) state to the 1′,4′-iminopyrimidine (IP) state. The identity of a general base was unknown which could not be easily determined using experimental techniques. A water-mediate mechanism (WMM) or direct histidine mechanism (DHM) mechanisms were proposed primarily comparison with the other enzymes sharing sequence and structural similarities to other TDP-dependent enzymes and transketolases, in particular. While the active sites of this family of enzymes have a high degree of similarity, the recent discovery of an enzyme deplete of acid/base residues in the active site and a transketolase lacking the requisite histidine provided impulse to investigate DXS further. A reaction coordinate, define as bond-breaking minus bond-forming, was used to incrementally change the system between the reactant and product states.

The WMM proposal was found to be preferential by a 11 kcal·mol⁻¹ difference in barrier heights. Computational results can be used in tandem with experimental results to help explain or reinforce conclusions made based on the experimental work. A H434A mutant revealed an effect on substrate binding while not effecting turnover. Thus, H434 was proposed not play a direct role in catalysis; which was supported in computationally via charge perturbation analysis. The charge of the H434 residue was artificially turned off and the result compared to the active site when the charge was on. The shift in the dipole resulting from this change illustrated the significance of this residue on long range electrostatics but no direct role in catalysis. Thus, the experimental work was bolstered and explained by the computational results.

Utilizing computational techniques, it is possible to investigate contributing factors otherwise inaccessible through experimental methods. The ΔE⁺_{WMM} was significantly higher than what would normally be expected for a simple proton transfer. A value of
22.7 kcal·mol$^{-1}$ for the WMM path vs 5-10 kcal·mol$^{-1}$ for representative proton transfers suggested some significant contributions. The deprotonation of TDP’s N4 atom produces a tautomerization from the AP state to the IP state, which we realized interrupts the aromaticity of the pyrimidinium ring. There is no method for directly computing the change in aromaticity in an enzyme reaction. There is a method for determining the aromaticity of the base TDP molecule and another for determining the percent change in aromaticity in reaction. By combining these methods, it was revealed by taking into account the change in aromaticity the $\Delta E^\ddagger_{WMM}$ would be closer to 8.5 kcal·mol$^{-1}$. The new value being closer to values previously published supported the conclusion of aromaticity playing a part in higher barrier energy. The higher energetic position of the IP state might also act as a driving force in the deprotonation of TDP’s C2 atom and production of the ylide required for TDP-dependent activity.

4.2 1-Deoxy-D-xylulose 5-Phosphate Reductoisomerase Summary and Conclusion

When the work began on 1-deoxy-D-xylulose 5-phosphate reductisomerase (DXR), there was much more of a debate in the literature over the last two mechanisms, $\alpha$-ketol rearrangement or retro-aldol/aldol mechanism. The reduction by NADPH has been pretty well understood since the enzymes initial characterizations. Along the way, papers were published with secondary kinetic isotope effects (KIEs) which supported the retro-aldol/aldol mechanism with the only remaining hope for the $\alpha$-ketol rearrangement was the refutation of the putative intermediates of the retro-aldol/aldol mechanism, namely hydroxyacetone enolate and glycoaldehyde phosphate. Since the lack of direct observation could be explained with a relatively simple assumption of being tightly bound in the active site requires less assumptions than trying to produce complicated assumptions to explain KIEs, Occam’s razor tentatively rules out the $\alpha$-ketol rearrangement in favor of the retro-aldol/aldol mechanism. So, the work was refocused on a new question pertaining to the binding of the Mg$^{2+}$ ion. Previously, it was thought that the metal bound
across the C2-C3 bond coordinated by their bound oxygens, but the same secondary KIEs shed light on the possibility of a second option. This option being bound across the C3-C4 bond via their oxygens. Besides explaining the observed KIEs, it would help to explain the lack of finding any intermediates since the metal would actually stretch over both intermediates instead of just one like in the C2-C3 mode.

The mechanistic work turned to helping to answer the binding mode question. Initial calculations attempted to utilize a single restraint as was done in the DXS mechanism. A single restraint was found wanting though. The results consistently produced highly strained configurations. In order to have better control, a second reaction coordinate was employed. The coordinates controlled related to the C3-C4 bond breaking and proton transfer from DXP O4 hydroxyl to a carboxyl atom of a glutamate residue 231. Since the first coordinate was a bond breakage, the $\delta_1$ was just stretched from 1.60Å to 3.60Å with 0.1Å increments. Similar to that of DXS, the proton transfer was a combination of the O4-H9 bond breaking and the H9-OE2 bond forming starting at the reactant state and progressing to the positive opposite value (i.e. -2.0Å to 2.0Å) over 0.2Å with the expectation the the final value would be shorter than the final 2.0Å mark.

The 2D-surfaces produced an ever increasing field for the C2-C3 binding mode while the C3-C4 binding mode had contours of interest for the retro-aldol reaction. The reaction consists of a deprotonation and C-C bond breakage producing the putative intermediates. Usually considered to be concerted, the C3-C4 energy surface has a valley at a point corresponding to the proton transfer progressing while the C-C breaking hadn’t begun. Proposing the possibility of a step-wise retro-aldol process might be a possibility. The C2-C3 surface lacks any signs suggesting this as a possible conclusion. Both of the final states were found to have comparatively high energies of 30.9 kcal·mol$^{-1}$ and 33.0 kcal·mol$^{-1}$ for the C3-C4 mode and C2-C3 mode, respectively. In addition to the energetics, the structure of the C2-C3 final state has a proton transferred to the phosphate tail of DXP instead of the D231 carboxyl group despite the restraint directing the other way. It is
possible the metal ion plays a role in preferential conformation stability and electron stabilization of the transition state through hydroxyl bond polarization.

Additional structural differences indicate a preference for the C3-C4 binding mode. Upon equilibration, this binding mode spreads across the C2-C3-C4 oxygens thus helping to coordinate the entire molecule. The additional binding would support the claims of tight binding in the active site throughout the reaction. The coordination promote the formation of the C2-C4 bond necessary the skeletal rearrangement. If this coordination is indeed necessary for aldolization, it might explain the lack of turnover when the hydroxyacetone and glycoaldehyde phosphate intermediate were exogenously introduced. The formation of the metal coordination might be very unlikely with the two intermediates compared to the single reactant.

4.3 Future Work

As previously mentioned, the projects above represent parts of a complete step of a reaction. The DXS work focused on the first half of ylide formation while the DXR project focuses on the retro-aldol reaction of the retro-aldol/aldol mechanism. Therefore, the follow up work should look to compute the completion of each step in the reaction. Additionally mapping the following reactions of DXS could provide valuable insight into residue contribution while the reactions are well understood. A project of interest would be the decarboxylation of pyruvate and subsequent transferral to G-3-P. Originally, the decarboxylation was thought to take place prior to G-3-P binding, but recent evidence suggests a pause until G-3-P binding. The energetics of decarboxylation with and without G-3-P and environmental analysis might provide unique insights into DXS.

These two enzymes represent the steps of the NMA pathway. Aspects the down-stream enzymes might be ascertained utilizing computational techniques while experimental methods don’t have the ability. The IspD enzyme is responsible for the transferral of the CMP group of CTP with a hide degree of specificity. Determining aspects to the specificity might be gleamed via Normal Mode Analysis coupled with Vibrational
Subsystem Analysis. Normal mode computes the frequencies of a system while vibration subsystem analysis determines how the large modes have on a subsystems of interest. Thus, mapping changes brought on via simulations might provide insight in the changes upon binding CTP; which in turn would provide further insights into residues of interest.

A mechanism keenly designed for computational investigation might be that the iron-sulfur cluster dependent IspG and IspH. Of particular interest would be IspH, the enzyme is able to produce by IDP and DMADP in a 4:1 ratio via a radical reaction. The ratio of 4:1 also represents the relative usage in downstream isoprenoids. An investigation into differences in the active site or energetics of the reaction might provide insight into how this enzyme preforms such an operation. There are many enzymes that have undesirable biproducts but no to my knowledge that produce both products of a pathway. The MVA pathway for instance utilizes isomerase to convert between IDP and DMADP.

NMA pathway enzymes are not found in mammalian cells suggest this pathway to be wonderful target to novel anti-biotic research. This is bolster by the fact that fosmidomycin is a known anti-malarial drug and inhibits DXR activity. There are other chemicals going through clinical trials currently with hopes of becoming a cheaper and better treatment for disease. Fosmidomycin has also been shown to inhibit IspD and IspE in addition to DXR though to a reduced extent. So it might even be possible to design a drug that target multiple enzymes, thus producing a stronger anti-microbial compound.
Bibliography


Appendix A: Supporting Information for “Thiamine Diphosphate Activation in 1-deoxy-D-xylulose 5-Phosphate Synthase: Insights into the Mechanism and Underlying Intermolecular Interactions”

A.1 Methods

A.1.1 Topology and Parameters for Thiamine Diphosphate (TDP)

TDP has never been parameterized for use in MM calculations. In order to build 1-deoxy-d-xylulose 5-phosphate synthase (DXS), it was necessary to develop a topology and parameter file that would reproduce the TDP crystal structure. TDP has a pyrimidine ring, thiazole ring, and an inorganic phosphate tail; each of which have been developed for use in CHARMM calculations. The topology and parameters for each moiety were used and augmented to account for TDP’s final structure. The additional bonds and parameters were determined based on structures with homologous chemical properties. QM calculations at the B3LYP/6-31G* level of theory were used to determine acquire initial charges for undefined atoms (e.g., C1 atom, vide infra). These charges were combined with those already established for each moiety. MacKerrell’s charge rules (mackerell.umaryland.edu/ff_dev.html) for substituents was followed to account for the linkers between groups. Finally, the atomic charges were balanced via manual manipulation and chemical intuition to equal the final -2.0 charge for TDP. A minimization of TDP was performed using the topology and parameter files to a tolerance of 0.002 kcal·mol⁻¹·Å⁻¹. The minimized structure’s RMSD was compared to that of the crystal
structure. The two structures were found to deviation by 0.1Å.

*Topology File for Thiamine Diphosphate using CGenFF Atom Types

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*Parameter File for Thiamine Diphosphate
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| CG324  | NG2R52 | CG2R51 | CG2R51 |   5.4000 | 2 | 180.00 |
| CG324  | NG2R52 | CG2R53 | SG2R50 |   6.0000 | 2 | 180.00 |
| CG2R51 | CG321  | CG321  | HGA2   |   0.1500 | 3 | 0.00   |
| CG2R51 | CG321  | CG321  | OG303  |   0.4000 | 3 | 180.00 |
| CG2R62 | CG324  | NG2R52 | CG2R53 |   0.2300 | 2 | 180.00 |
| CG2R62 | CG324  | NG2R52 | CG2R51 |   0.2300 | 2 | 180.00 |
| CG2R51 | NG2R52 | CG324  | HGA2   |   0.0000 | 3 | 0.00   |
| CG2R53 | NG2R52 | CG324  | HGA2   |   0.0000 | 3 | 0.00   |
| CG2R62 | CG2R62 | CG324  | HGA2   |   0.0000 | 3 | 0.00   |
| CG2R64 | CG2R62 | CG324  | HGA2   |   0.0000 | 2 | 0.00   |
| NG2R52 | CG2R51 | CG331  | HGA3   |   0.1900 | 3 | 0.00   |
| SG2R50 | CG2R51 | CG321  | HGA2   |   0.1900 | 3 | 0.00   |
| CG2R51 | CG2R51 | CG321  | CG321  |   0.2000 | 1 | 0.00   |
| CG2R51 | CG2R51 | CG321  | CG321  |   0.2700 | 2 | 0.00   |
| CG2R51 | CG2R51 | CG321  | CG321  |   0.0000 | 3 | 0.00   |
| SG2R50 | CG2R51 | CG321  | CG321  |   0.1900 | 3 | 0.00   |
| CG2R62 | CG2R62 | CG324  | NG2R52 |   0.1500 | 2 | 180.00 |
| CG2R64 | CG2R62 | CG324  | NG2R52 |   0.1500 | 2 | 180.00 |
| CG2R51 | NG2R52 | CG2R53 | HGR52  |   2.0000 | 2 | 180.00 |
| CG2R64 | NG2R62 | CG2R62 | HGR62  |   4.5000 | 2 | 180.00 |
| CG2R53 | NG2R52 | CG2R51 | CG331  |   3.0000 | 2 | 180.00 |
| CG2R53 | SG2R50 | CG2R51 | CG321  |   8.5000 | 2 | 180.00 |
A.1.2 Extended 20 ns Molecular Dynamics Simulation

The crystal structure for the *D. radiodurans* DXS (PDB ID:201X) enzyme with TDP bound was processed and parsed via www.charmming.org. The topology and parameters for TDP shown above were used in generating the structure used in this extended simulation. Structural modifications were performed to ensure the active site Glu373 was protonated in agreement with experimental evidence. CGenFF and CHARMM22 protein (C22) force fields were used throughout. The system was solvated in a cubic crystal structure and neutralized with KCl salt to a final concentration of 0.15M. The system was heated from 110K to 310K over 100ps. The system was simulated for 20 ns at constant pressure (1atm) and temperature (310K). For the purposes of analysis, the first 2 ns were discarded to allow for an equilibration period.
A.1.3 Over-expression and Purification of Wildtype DXS and the DXS Mutants

Plasmids containing the wild type D. radiodurans dxs gene or the mutant dxs gene were transformed into E. coli BL-21 B(DE3) cells and used for protein expression. An overnight culture of E. coli in LB broth containing 50 µg/ml kanamycin was diluted 100-fold, cultured at 37°C until the absorbance at 600 nm reached ~ 0.6, and then cooled to 20°C. Expression was induced by the addition of 0.5 mM isopropyl β-D-1-thiogalactopyranoside (IPTG). The cells were harvested by centrifugation (6,000 × g for 10 min) after being shaken for 6 hrs at 20°C and the resulting cell pellets stored at -80°C before purification. Cells were thawed and all the purification steps were performed at 4°C. Cells were resuspended in binding buffer (20 mM Tris, 500 mM NaCl, 5 mM imidazole, 10 mM β-Me, pH = 7.5) supplemented with 1 mM phenylmethanesulfonylfluoride (PMSF), 4 µg/mL leupeptin, and 2 µg/mL pepstatin, sonicated using a Heat systems W-380 ultrasonic processor, and centrifuged (16,000 × g for 20 min) to remove cell debris. The supernatant from the cell lysate was applied to a 1.5 cm × 5 cm column packed with Ni-NTA resin that had been equilibrated with binding buffer. Non-bound proteins eluted from the column by first washing with 5 column volumes of binding buffer followed by 20 column volumes of wash buffer (20 mM Tris pH 7.5, 500 mM NaCl, 60 mM imidazole, and 10 mM β-Me). The bound DXS (wildtype or mutant) was eluted from the Ni-NTA resin using elution buffer (20 mM Tris pH 7.5, 500 mM NaCl, 250 mM imidazole, and 10 mM β-Me). A flow rate of 1.5 mL/min was maintained through the Ni-NTA column for all the loading and washing steps. DXS-containing fractions containing were combined, exhaustively dialyzed at 4°C against 20 mM Tris pH 7.5, 100 mM NaCl, and 10 mM β-Me, and concentrated by ultrafiltration. The final yield of DXS (wildtype or mutant) was 7-8 mg/L of E. coli culture medium. Enzyme was flash frozen in liquid nitrogen, stored at -80°C. The purity of the DXS (wildtype or the mutant) was evaluated by SDS-PAGE.
Figure A.1: An illustration of the QM regions of both the direct histidine mechanism (DHM, image (a)) and the water-mediated mechanism (WMM, image (b)). DHM’s QM region contained 98 atoms; which were made up from H124, E373, H434 and TDP. WMM’s QM region was comprised of all the same residues as the DHM with the addition of Wat9709 (reactive water) making the total 101 atoms. Additionally, linker atoms were used the C-O-Cα bond (purple atoms).
Figure A.2: Plots of the initial velocities versus varying concentrations of pyruvate or G3P. Plots a and b are for wild-type DXS while plots c and d represent the H82A mutant.
Figure A.3: Graphs of initial velocities versus varying concentrations of pyruvate or G3P. Plots a and b are for the H304A mutant. Plots c and d represent the D430A mutant.
Figure A.4: Plots of the initial velocities versus varying concentrations of pyruvate or G3P. All graphs are for the H434A mutant discussed in primary manuscript.
A.2 ProPKA3.1 Results

Below will be found the ProPKA3.1 results for monomer A of DXS:
Figure A.5: This bar graph represents all of the CPA data acquired for all of the residues within 5.0 Å of the QM region. DHM is in gold while WMM is in green. What is immediately apparent is the significantly larger change in magnitude for all DHM results in comparison to that of the WMM. This is thought to come from the large change in the active site dipole.
References:

Very Fast Empirical Prediction and Rationalization of Protein pKa Values
Hui Li, Andrew D. Robertson and Jan H. Jensen

Very Fast Prediction and Rationalization of pKa Values for Protein-Ligand Complexes
Delphine C. Bas, David M. Rogers and Jan H. Jensen

PROPKA3: Consistent Treatment of Internal and Surface Residues in Empirical pKa predictions
Mats H.M. Olsson, Chresten R. Sondergard, Michal Rostkowski, and Jan H. Jensen

Improved Treatment of Ligands and Coupling Effects in Empirical Calculation
and Rationalization of pKa Values
Chresten R. Sondergaard, Mats H.M. Olsson, Michal Rostkowski, and Jan H. Jensen
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ASP 154 A 0.28 ASP 182 A -0.23 ASN 183 A -0.11 HIS 284 A
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ASP 610 A 4.24 17 % 0.89 329 0.04 0 -0.03 ASP 610 A 0.01 GLU 601 A
ASP 610 A 0.00 XXX 0 X -0.35 ALA 613 A -0.12 HIS 604 A
ASP 624 A 3.93 0 % 0.13 161 0.00 0 -0.88 ARG 477 A
GLU 28 A 4.41 0 % 0.26 185 0.00 0 -0.83 LEU 12 A -0.03 ARG 43 A
GLU 35 A 4.59 0 % 0.16 277 0.00 0 -0.96 ARG 38 A
GLU 35 A 0.00 XXX 0 X -0.00 LYS 291 A
GLU 36 A 4.95 28 % 1.84 359 0.22 0 -0.83 LEU 13 A -0.11 ARG 94 A
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GLU 40 A -0.69 ARG 94 A 0.08 ASP 95 A
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GLU 103 A 3.66 0 % 0.37 248 0.00 0 -0.85 SER 602 A 0.00 XXX 0 X -0.33 ARG 606 A
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GLU 114 A 4.03 64 % 2.45 461 0.71 0 -0.85 LYS 88 A -0.80 SER 107 A -0.09 LYS 111 A
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GLU 114 A 0.00 XXX 0 X -0.18 HIS 597 A
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GLU 116 A 4.36 0 % 0.31 255 0.00 0 0.00 XXX 0 X 0.00 XXX 0 X -0.07 LYS 20 A

http://propka.ki.ku.dk/pka/2o1x.pka
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GLU 116 A 0.00 XXX 0 X 0.00 XXX 0 X -0.01 ARG 75 A
GLU 116 A 0.00 XXX 0 X 0.00 XXX 0 X -0.25 HIS 117 A

GLU 184 A 5.15 16 % 0.32 325 0.04 0 0.00 XXX 0 X 0.00 XXX 0 X -0.08 MG MG A
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GLU 189 A 3.74 29 % 0.67 362 0.00 0 -0.33 ARG 350 A -0.61 GLU 189 A -0.03 LYS 289 A
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GLU 189 A 0.00 XXX 0 X 0.00 XXX 0 X 0.03 ASP 299 A
GLU 189 A 0.00 XXX 0 X 0.00 XXX 0 X 0.43 ARG 350 A

GLU 266 A 5.09 7 % 0.43 300 0.05 0 0.00 XXX 0 X 0.00 XXX 0 X 0.11 ASP 260 A
GLU 272 A 4.24 8 % 0.62 304 0.05 0 -0.54 ARG 27 A 0.00 XXX 0 X -0.40 ARG 27 A

GLU 297 A 4.13 17 % 0.70 330 0.13 0 0.00 XXX 0 X -0.64 GLY 292 A -0.10 ARG 38 A
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GLU 315 A 4.73 0 % 0.12 142 0.00 0 0.00 XXX 0 X 0.00 XXX 0 X 0.11 ASP 310 A

GLU 330 A 4.67 20 % 0.90 337 0.12 0 -0.14 GLN 485 A 0.00 XXX 0 X -0.01 LYS 337 A
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GLU 330 A 0.00 XXX 0 X 0.00 XXX 0 X 0.05 GLU 357 A

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GLU 373 A 8.47 100 % 3.73 588 0.84 0 0.00 XXX 0 X -0.53 GLU 373 A 0.57 GLU 374 A
GLU 373 A 0.00 XXX 0 X 0.00 XXX 0 X -0.64 ARG 401 A
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ARG 401 A
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ARG 423 A
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ARG 444 A
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ARG 501 A
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ARG 536 A
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ARG 536 A
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ARG 586 A 12.70  0 %  -0.36  281  0.00  0  0.22 GLU 620 A  0.00 XXX  0 X  0.34 GLU 620 A
ARG 606 A 12.40  3 %  -0.44  291  0.00  0  0.00 XXX  0 X  0.00 XXX  0 X  0.33 GLU 103 A
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TDP O21 A -0.86* 100 %  3.86  613  0.00  0  -0.85 SER 54 A  0.00 XXX  0 X  -3.35 MG  MG A
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TDP O21 A  -0.17 HIS 304 A  0.00 XXX  0 X  -2.03 LYS 289 A
TDP O21 A  0.00 XXX  0 X  0.00 XXX  0 X  -1.46 HIS 304 A

Coupled residues (marked *) were detected. Please rerun PropKa with the --display-coupled-residues
or -d option for detailed information.

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Free energy of folding (kcal/mol) as a function of pH (using neutral reference)

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The pH of optimum stability is 6.7 for which the free energy is 45.5 kcal/mol at 298K

Could not determine pH values where the free energy is within 80% of minimum
Could not determine the pH-range where the free energy is negative

Protein charge of folded and unfolded state as a function of pH

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The pI is 6.00 (folded) and 6.00 (unfolded)
Below will be found the ProPKA3.1 results for monomer B of DXS:
References:

Very Fast Empirical Prediction and Rationalization of Protein pKa Values
Hui Li, Andrew D. Robertson and Jan H. Jensen

Very Fast Prediction and Rationalization of pKa Values for Protein-Ligand Complexes
Delphine C. Bas, David M. Rogers and Jan H. Jensen

PROPKA3: Consistent Treatment of Internal and Surface Residues in Empirical pKa predictions
Mats H.M. Olsson, Chresten R. Sondergard, Michal Rostkowski, and Jan H. Jensen

Improved Treatment of Ligands and Coupling Effects in Empirical Calculation and Rationalization of pKa Values
Chresten R. Sondergard, Mats H.M. Olsson, Michal Rostkowski, and Jan H. Jensen
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ARG 552 B 12.73 0 % -0.23 222 0.00 0 0.00 XXX 0 X 0.00 XXX 0 X 0.18 ASP 506 B
ARG 552 B 0.00 XXX 0 X 0.00 XXX 0 X 0.28 GLU 548 B
ARG 554 B 13.25 0 % -0.35 266 0.00 0 0.71 ASP 507 B 0.00 XXX 0 X 0.01 ASP 624 B
ARG 554 B 0.00 XXX 0 X 0.00 XXX 0 X 0.38 ASP 507 B
ARG 586 B 12.35 0 % -0.28 261 0.00 0 0.00 XXX 0 X 0.00 XXX 0 X 0.14 GLU 620 B
ARG 606 B 12.38 3 % -0.47 291 0.00 0 0.00 XXX 0 X 0.00 XXX 0 X 0.35 GLU 103 B
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ARG 615 B 12.80 0 % -0.46 279 0.00 0 0.22 GLU 628 B 0.00 XXX 0 X 0.07 GLU 525 B
ARG 615 B 0.00 XXX 0 X 0.00 XXX 0 X 0.38 GLU 628 B
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N+ 8 B 0.00 XXX 0 X 0.00 XXX 0 X 0.14 ASP 14 B
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TDP O21 B 0.00 XXX 0 X 0.00 XXX 0 X -1.12 HIS 304 B

Coupled residues (marked *) were detected. Please rerun PropKa with the --display-coupled-residues or -d option for detailed information.

SUMMARY OF THIS PREDICTION
   Group   pKa  model-pKa  ligand atom-type
     ASP    9 B  3.68    3.80

http://propka.ki.ku.dk/pka/2o1x.pka
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Free energy of folding (kcal/mol) as a function of pH (using neutral reference)

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The pH of optimum stability is 6.8 for which the free energy is 47.4 kcal/mol at 298K. Could not determine pH values where the free energy is within 80% of minimum. Could not determine the pH-range where the free energy is negative.

Protein charge of folded and unfolded state as a function of pH:

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The pI is 5.40 (folded) and 5.80 (unfolded).
Appendix B: Supporting Information for “Computational Examination of the Magnesium Ion Binding Modes of 1-Deoxy-D-xylulose 5-Phosphate Reductoisomerase”

B.1 Results from the ProPKA3.0 Calculations of a DXR with Substrates Bound

Below will be found the ProPKA3.0 results for of crystal structure, PDB:1Q0Q, for DXR with bound NADPH and DXP:
# propka3.0, revision 182

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**PROPKA: A PROTEIN PKA PREDICTOR**

---

**VERSION 1.0, 04/25/2004, IOWA CITY**

**BY HUI LI**

---

**VERSION 2.0, 11/05/2007, IOWA CITY/COPENHAGEN**

**BY DELPHINE C. BAS AND DAVID M. ROGERS**

---

**VERSION 3.0, xx/xx/2010, COPENHAGEN**

**BY MATS H.M. OLSSON AND CHERSTEN R. SONDERGARD**

---

**References:**

- **Very Fast Empirical Prediction and Rationalization of Protein pKa Values**
  Hui Li, Andrew D. Robertson and Jan H. Jensen

- **Very Fast Prediction and Rationalization of pKa Values for Protein-Ligand Complexes**
  Delphine C. Bas, David M. Rogers and Jan H. Jensen

- **PROPKA3: Consistent Treatment of Internal and Surface Residues in Empirical pKa predictions**
  Mats H.M. Olsson, Chresten R. Sonderegard, Michal Rostkowski, and Jan H. Jensen

---

**WARNING!**

Propka3.0 is not identical to propka2.0 and does not work with ligands

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GLU 231 B  -0.85 LYS 228 B  0.00 XXX  0 X  0.00 XXX  0 X
GLU 234 B  12.78*  100 %  4.37  712  0.66  0  1.21 ASP 150 B  0.00 XXX  0 X  -1.14 LYS 125 B
GLU 234 B  0.00 XXX  0 X  0.00 XXX  0 X  -0.06 LYS 228 B
GLU 234 B  0.00 XXX  0 X  0.00 XXX  0 X  0.18 GLU 126 B
GLU 234 B  0.00 XXX  0 X  0.00 XXX  0 X  0.34 GLU 152 B
GLU 234 B  0.00 XXX  0 X  0.00 XXX  0 X  1.97 ASP 150 B
GLU 234 B  0.00 XXX  0 X  0.00 XXX  0 X  0.78 GLU 231 B
GLU 247 B  3.30  58 %  1.63  445  0.04  0  -0.80 SER 180 B  0.00 XXX  0 X  -0.80 ARG 261 B
GLU 247 B  -1.27 ARG 261 B  0.00 XXX  0 X  0.00 XXX  0 X
GLU 273 B  4.91  28 %  0.85  361  0.10  0  -0.29 LYS 295 B  0.00 XXX  0 X  -0.03 ARG 261 A
GLU 273 B  0.00 XXX  0 X  0.00 XXX  0 X  0.09 ASP 275 B
GLU 273 B  0.00 XXX  0 X  0.00 XXX  0 X  0.05 ASP 298 B
GLU 273 B  0.00 XXX  0 X  0.00 XXX  0 X  -0.36 LYS 295 B
GLU 323 B  4.58  0 %  0.34  282  0.00  0  0.00 XXX  0 X  0.00 XXX  0 X  -0.27 LYS 319 B
GLU 326 B  4.71  0 %  0.16  216  0.00  0  0.00 XXX  0 X  0.00 XXX  0 X  -0.05 ARG 236 B
GLU 326 B  0.00 XXX  0 X  0.00 XXX  0 X  0.10 GLU 323 B
GLU 340 B  3.93  100 %  2.89  577  0.65  0  -0.48 ASN 336 B  -0.75 ALA 104 B  -1.03 LYS 217 B
GLU 340 B  -0.52 ARG 386 B  0.00 XXX  0 X  -1.33 ARG 386 B
GLU 365 B  4.68  0 %  0.26  223  0.00  0  0.00 XXX  0 X  0.00 XXX  0 X  -0.10 LYS 366 B
GLU 365 B  0.00 XXX  0 X  0.00 XXX  0 X  0.02 GLU 392 B
GLU 371 B  3.85  18 %  0.81  333  0.08  0  0.00 XXX  0 X  -0.68 GLN 329 B  -0.14 ARG 370 B
GLU 371 B  0.00 XXX  0 X  -0.71 ALA 330 B  0.00 XXX  0 X
GLU 387 B  3.95  0 %  0.22  204  0.00  0  -0.20 ARG 390 B  0.00 XXX  0 X  -0.26 ARG 390 B
GLU 387 B  0.00 XXX  0 X  0.00 XXX  0 X  -0.31 LYS 391 B
GLU 392 B  2.80  1 %  0.53  283  0.00  0  -0.85 SER 362 B  0.00 XXX  0 X  -0.00 LYS 391 B
GLU 392 B  -0.69 LYS 366 B  0.00 XXX  0 X  -0.31 ARG 395 B
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C-  398 B  3.30  0 %  0.10  110  0.00  0  0.00 XXX  0 X  0.00 XXX  0 X  0.00 XXX  0 X
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Residues that are found to be 'coupled', i.e. titrates together, has been marked by '*' in the above section. Please rerun PropKa with the --display-coupled-residues option for detailed information.

SUMMARY OF THIS PREDICTION

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</tr>
<tr>
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</tr>
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<tr>
<td>ARG</td>
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</table>
### Free energy of folding (kcal/mol) as a function of pH (using neutral reference)

<table>
<thead>
<tr>
<th>pH</th>
<th>unfolded</th>
<th>folded</th>
</tr>
</thead>
<tbody>
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<td>115.93</td>
<td>89.99</td>
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<tr>
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<tr>
<td>4.00</td>
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<tr>
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</table>

The pH of optimum stability is 6.7 for which the free energy is 69.2 kcal/mol at 298K.

Could not determine pH values where the free energy is within 80% of maximum.

Could not determine where the free energy is positive.

### Protein charge of folded and unfolded state as a function of pH

<table>
<thead>
<tr>
<th>pH</th>
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<th>folded</th>
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</thead>
<tbody>
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</tr>
</tbody>
</table>

The pI is 5.65 (folded) and 5.97 (unfolded).