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Factors Contributing to the Biofilm-Deficient Phenotype of *Staphylococcus aureus* sarA Mutants

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**Abstract**

Mutation of *sarA* in *Staphylococcus aureus* results in a reduced capacity to form a biofilm, but the mechanistic basis for this remains unknown. Previous transcriptional profiling experiments identified a number of genes that are differentially expressed both in a biofilm and in a *sarA* mutant. This included genes involved in acid tolerance and the production of nucleolytic and proteolytic exoenzymes. Based on this we generated mutations in *alsSD*, *nuc* and *spa* in the *S. aureus* clinical isolate UAMS-1 and its isogenic *sarA* mutant and assessed the impact on biofilm formation. Because expression of *alsSD* was increased in a biofilm but decreased in a *sarA* mutant, we also generated a plasmid construct that allowed expression of *alsSD* in a *sarA* mutant. Mutation of *alsSD* limited biofilm formation, but not to the degree observed with the corresponding *sarA* mutant, and restoration of *alsSD* expression did not restore the ability to form a biofilm. In contrast, concomitant mutation of *sarA* and *nuc* significantly enhanced biofilm formation by comparison to the *sarA* mutant. Although mutation of *spa* had no significant impact on the ability of a *sarA* mutant to form a biofilm, a combination of protease inhibitors (E-64, 1-10-phenanthroline, and dichloroisoucumin) that was shown to inhibit the production of multiple extracellular proteases without inhibiting growth was also shown to enhance the ability of a *sarA* mutant to form a biofilm. This effect was evident only when all three inhibitors were used concurrently. This suggests that the reduced capacity of a *sarA* mutant to form a biofilm involves extracellular proteases of all three classes (serine, cysteine and metalloproteases). Inclusion of protease inhibitors also enhanced biofilm formation in a *sarA/nuc* mutant, with the combined effect of mutating *nuc* and adding protease inhibitors resulting in a level of biofilm formation with the *sarA* mutant that approached that of the UAMS-1 parent strain. These results demonstrate that the inability of a *sarA* mutant to repress production of extracellular nucleases and multiple proteases have independent but cumulative effects that make a significant contribution to the biofilm-deficient phenotype of an *S. aureus* *sarA* mutant.

**Introduction**

*Staphylococcus aureus* is an opportunistic pathogen capable of causing diverse forms of infection. Treatment of these infections is complicated not only by the continued emergence of antibiotic-resistant strains but also by the fact that many *S. aureus* infections are associated with formation of a biofilm, which limits the efficacy of antimicrobial therapy even in cases caused by strains that are not clinically defined as resistant to the relevant antibiotics [1,2]. For this reason, the effective treatment of biofilm-associated staphylococcal infections often requires surgical debridement to remove infected tissues and/or devices [3,4].

Previous reports have implicated many *S. aureus* genes in biofilm formation. These include *agr* [5,6], *arrRS* [7], *bam* [8], *hla* [9], *ica* [10], *rfb* [11], *sacA* [12,13], *sigB* [14,15], *teaR* [16], and *traP* [17,18]. However, in almost all cases, there are conflicting reports with respect to the overall contribution of specific loci. For example, some reports have concluded that *tea* and *sigB* are required for *S. aureus* biofilm formation [10,14,15], while others have found that mutation of these loci has little impact [13,19,20]. Similarly, there is a report concluding that alpha hemolysin is required for biofilm formation [9], but *S. aureus* isolates unable to produce alpha toxin owing to a nonsense mutation in the corresponding gene (*hla*) are capable of forming a biofilm and causing biofilm-associated infection [19,21]. In fact, a recent report concluded that non-hemolytic variants arise spontaneously within a biofilm and ultimately become the dominant subpopulation [22].

Such conflicting reports may be due to strain-dependent differences among *S. aureus* isolates. For instance, *bap* encodes a surface-associated protein (Bap) that promotes biofilm formation, but to date it has been found only in bovine mastitis isolates and even then only rarely [23,24]. Many studies focusing on biofilm formation have also utilized strains derived from NCTC 8325. This includes RN6390, which is an 8325-4 strain in which three prophage were cured from NCTC 8325 [25], and SA113, which is a mutated, restriction-modification deficient derivative of 8325 [10,26]. All 8325-derived strains carry natural mutations in *rfbU*, which renders them functionally *sigB* deficient [27–29]. They also carry a mutation in *kaF* [16], a regulatory locus that has also been implicated in biofilm formation [30]. Perhaps owing to these
mutations, the 8325-4 strain RN6390 has a reduced capacity to form a biofilm by comparison to clinical isolates of *S. aureus* [19,21].

One exception to such conflicting reports is the staphylococcal accessory regulator (sarA), mutation of which has been consistently correlated with a reduced capacity to form a biofilm in both *S. aureus* and *S. epidermidis* [12,13,21]. Indeed, with the exceptions of RN6390 and Newman, the latter also being a poor biofilm former [12] that has specific characteristics (e.g. production of truncated fibronectin-binding proteins) that distinguish it from primary clinical isolates [31], mutation of *sarA* has resulted in a reduced capacity to form a biofilm *in vivo* in every *S. aureus* strain we have examined [12].

Mutation of *sarA* in the clinical isolate UAMS-1 was also shown to result in a significant decrease in biofilm formation *in vivo* as defined using a catheter-based murine model [19].

The *sarC* locus encodes a DNA-binding protein (SarA) that has a global impact on gene expression in *S. aureus* [21,32], and it remains unclear which components of this global response are most relevant to biofilm formation. Mutation of *sarC* does result in reduced expression of the *ica* operon and consequently reduced production of the polysaccharide intercellular adhesion (PIA), but our direct comparison of *sarA* and *ica* mutants generated in the same strain used in the experiments reported here (UAMS-1) demonstrate that this cannot account for the biofilm defect in a *sarA* mutant [19].

As a first step toward defining the role of *sarA* in biofilm formation, we compared the regulons defined by growth within a biofilm and by mutation of *sarA*. We found that a large number of genes were differentially expressed in a mature biofilm by comparison to both exponential and post-exponential phase planktonic cultures [19] and that many of these were also in the *sarA* regulon [33]. Included among these genes was the bicistronic operon *alsSD*, which encodes the enzymes (acetolactate synthase and acetolactate decarboxylase) required for the conversion of pyruvate to acetoin rather than the more acidic products of glucose metabolism. Specifically, expression of *alsSD* was increased in a biofilm by comparison to both exponential and post-exponential planktonic growth [12]. Conversely, *alsSD* expression was decreased in a *sarA* mutant [33]. This suggests that the inability to express *alsSD* at adequate levels in a *sarA* mutant may contribute to its inability to form a biofilm.

Also included in both the biofilm and *sarA* regulons was *nuc*, which encodes the *S. aureus* thermostable nuclease. The expression pattern of *nuc* was opposite that of *alsSD* in that it was decreased in a biofilm but increased in a *sarA* mutant [19,33]. This observation, together with recent reports demonstrating that extracellular DNA (eDNA) contributes to *S. aureus* biofilm formation [34,35], suggest that the inability to repress production of extracellular nucleases may also contribute to the biofilm-deficient phenotype of a *sarA* mutant.

Expression of the genes encoding extracellular proteases was also altered both in a biofilm and a *sarA* mutant, but in this case the scenario is more complex in that expression of some protease genes (e.g. *scpA*) was decreased in a biofilm while expression of others (e.g. *spaB*) was increased [19]. However, expression of all of these genes, as well as the gene encoding aureolysin (*aur*) was increased in a *sarA* mutant [33]. This is consistent with the observation that the overall production of extracellular proteases is increased in a *sarA* mutant [36–38]. This is potentially important in that biofilm formation in clinical isolates of *S. aureus* is facilitated by coating the substrate with plasma proteins [12], and the increased protease production observed in *sarA* mutants has been correlated with a reduced capacity to bind host proteins including fibronectin [36,39]. Moreover, a recent report concluded that the *agr*-mediated induction of protease production limits biofilm formation and may play a functional role with respect to the dispersal of *S. aureus* from an established biofilm [40]. Such results suggest that the reduced capacity of a *sarC* mutant to form a biofilm could also be related to the increased production of one or more extracellular proteases.

Based on these considerations, we investigated the role of *alsSD*, *nuc*, and extracellular proteases in *S. aureus* biofilm formation with a specific emphasis on whether any or all of these factors contribute to the biofilm-deficient phenotype of a *sarA* mutant.

**Results And Discussion**

**Impact of *sarA* on *alsSD* expression, acetoin production, stationary phase survival and biofilm formation**

The *alsS* and *alsD* genes constitute a bicistronic operon and encode acetolactate synthase and acetolactate decarboxylase, respectively. These enzymes function sequentially to convert pyruvate to 2-acetolactate and then acetoin, the latter ultimately being converted by acetoin reductase to 2,3-butanediol. Production of these neutral products rather than the more acidic products of carbohydrate metabolism is important for acid tolerance in a number of bacterial species [41]. This is consistent with the observation that *alsSD* is expressed at elevated levels in response to mild acid treatment of *S. aureus* [42]. Our previous demonstration that expression of *alsSD* is also increased in a biofilm by comparison to planktonic cultures is therefore consistent with the hypothesis that a central theme behind the adaptation of *S. aureus* to persistence within a biofilm is survival within the acidic environment associated with carbohydrate metabolism, particularly under low oxygen conditions [19]. It is also consistent with a recent report demonstrating that *Streptococcus mutans* is more acid tolerant when grown in a biofilm than when grown in planktonic culture [43]. Moreover, production of the enzymes required for acetoin production in *Vibrio cholerae* is co-regulated with expression of genes directly involved in the switch between motility and biofilm formation [41]. This co-regulation provides further support for the hypothesis that acetoin production and acid tolerance are important for biofilm formation in diverse bacterial species.

A previous report described construction of a UAMS-1 *alsSD* mutant (KB1097) and concluded that mutation of *alsSD* eliminated acetoin production and resulted in reduced murein hydrolase activity and reduced stationary phase survival [44]. We subsequently demonstrated that KB1097 also has a reduced capacity to form a biofilm that is comparable to that observed with a UAMS-1 *sarA* mutant [33]. However, it was later discovered that KB1097 was a contaminant that ultimately proved to be *S. hominis* rather than *S. aureus* (Dr. Ken Bayles, personal communication). Based on this, we independently generated an *alsSD* mutant (UAMS-1489) using the pKOR1 mutagenesis system [45]. Characterization of this mutant confirmed the absence of *alsSD* transcription (Fig. 1), the inability to produce acetoin and 2,3-butanediol as defined by the Voges-Proskauer assay (Fig. 2), and reduced stationary-phase survival (Fig. 3). The results also confirmed that mutation of *sarA* impacts all three phenotypes in a similar although less definitive manner (Figs. 1–3). Additionally, the capacity to form a biofilm was significantly reduced in UAMS-1489 by comparison to the UAMS-1 parent strain (p<0.001) (Fig. 4). Transcription of *alsSD*, the Voges-Proskauer phenotype, stationary-phase survival, and biofilm formation were all complemented by introducing a functional *alsSD* operon into UAMS-1489 to generate UAMS-1551 (Figs. 1–4). These results are consistent with the hypothesis that *alsSD* plays a role in biofilm formation in *S. aureus*.

While UAMS-1489 had a reduced capacity to form a biofilm, it remained significantly greater than that observed with the UAMS-929 *sarA* mutant (p<0.001) (Fig. 4). This was true despite the fact...
that mutation of alsSD had a greater impact than mutation of sarA on alsSD transcription, the Voges-Proskauer assay, and stationary-phase survival (Fig. 1–4). This demonstrates that the impact of sarA on biofilm formation extends beyond its impact on transcription of alsSD. Confirmation of this was obtained by analysis of the sarA/alsSD double mutant (UAMS-1300), which demonstrated that concomitant mutation of alsSD and sarA had no further impact on the production of acetoin (data not shown) but did reduce biofilm formation even by comparison to an alsSD mutant (p<0.001) (Fig. 4).

Although these results demonstrate that the impact of sarA on alsSD transcription cannot fully account for the biofilm-deficient phenotype of a sarA mutant, mutation of alsSD did have a significant impact on biofilm formation by comparison to the parent strain (Fig. 4), and this leaves open the possibility that the impact of sarA on alsSD transcription makes an important contribution in that regard. Based on this, we also generated derivatives of UAMS-929 (UAMS-1729 and UAMS-1730) in which alsSD expression was enhanced in a sarA mutant. Although this did not fully restore alsSD transcription to wild-type levels

Figure 1. Expression of alsSD in alsSD and sarA mutants. The relative level of alsSD expression in each strain was determined by qRT-PCR. Results represent the mean±standard deviation of 3 replicate samples. Strain designations are: U1, UAMS-1 (parent strain); U929, UAMS-1 sarA mutant; U969, complemented sarA mutant; U1489, UAMS-1 alsSD mutant; U1551, complemented alsSD mutant. doi:10.1371/journal.pone.0003361.g001

Figure 2. Production of acetoin and 2,3-butanediols in alsSD and sarA mutants. 18-hour culture supernatants were assayed for acetoin production using the Voges-Proskauer assay. Strain designations are the same as those cited in Fig. 1 and detailed in Table 1. Results represent the mean±standard deviation of 6 replicate experiments. doi:10.1371/journal.pone.0003361.g002
(Fig. 5A), which might be expected given the positive impact of sarA on alsSD transcription [33], it did restore transcription to levels sufficient to fully restore the Voges-Proskauer phenotype (Fig. 5B). However, it had no impact on the ability of a sarA mutant to form a biofilm (Fig. 5C). This provides further support for the conclusion that mutation of sarA results in a defect in biofilm formation that extends beyond its impact on expression of alsSD.

One possible explanation for these results is that alsSD is part of a pathway that includes acetoin reductase, which is required to convert acetoin to 2,3-butanediol, and expression of the corresponding gene (SA0239 in the N315 genome) was also reduced in a sarA mutant [33]. Thus, restoration of acetoin production in a sarA mutant may not fully restore the ability to convert pyruvate to 2,3-butanediol. While such an effect might not be evident in the Voges-Proskauer assay, it could nevertheless compromise other aspects of metabolism including the ability to maintain pH homeostasis. However, subsequent analysis of all relevant strains failed to reveal any significant difference in the pH of culture supernatants from overnight cultures grown in biofilm medium, which includes exogenous glucose (data not shown). The alternative explanation is that sarA has an impact on biofilm formation that involves genes unrelated to the acetoin/2,3-butanediol pathway.

**Contribution of thermostable nuclease to the biofilm-deficient phenotype of a sarA mutant**

Previous genome-scale transcriptional profiling experiments focusing on UAMS-1 identified a number of other sarA-regulated genes that were also differentially expressed in a biofilm by comparison to planktonic cultures [19,33]. Included among these genes was nuc, which encodes the *S. aureus* thermostable nuclease. Specifically, expression of nuc was reduced in a biofilm and increased in a sarA mutant [12,33]. Recent reports demonstrating that extracellular DNA contributes to biofilm formation in *S. aureus*
[34,35] are consistent with the hypothesis that this may also contribute to the biofilm-deficient phenotype of a sarA mutant.

To address this hypothesis, we also generated UAMS-1 nuc and sarA/nuc mutants and evaluated the impact on biofilm formation. Phenotypic assays confirmed that mutation of sarA increased nuclease production and that it was completely abolished by mutation of nuc even in a sarA mutant (Fig. 6). Mutation of nuc in UAMS-1 (UAMS-1471) had no impact on biofilm formation, but complementation of the nuc mutation with a plasmid-borne version of the nuc gene (UAMS-1552) did limit biofilm formation to a significant degree \( p = 0.001 \) (Fig. 7), presumably because it resulted in increased production of nuclease by comparison to the parent strain (Fig. 6). More importantly, concomitant mutation of nuc in a sarA mutant (UAMS-1477) enhanced biofilm formation by comparison to the sarA mutant \( p < 0.001 \) (Fig. 7). This effect was completely reversed by complementation of the sarA/nuc double mutant with a plasmid-borne version of the nuc gene (UAMS-1725) (Fig. 7). At the same time, biofilm formation in the sarA/nuc mutant (UAMS-1477) was not restored to wild-type levels. In contrast, complementation of the sarA/nuc mutant with sarA did fully restore biofilm formation (Fig. 7). These results indicate that increased production of extracellular nuclease contributes to the biofilm-deficient phenotype of a UAMS-1 sarA mutant but that additional factors must also be involved.

**Contribution of extracellular proteases to the biofilm-deficient phenotype of a sarA mutant**

Also included in both the biofilm and sarA regulons were most of the S. aureus genes encoding extracellular proteases, the only exception being the spl operon \( \text{splABCDEF} \), expression of which was largely unaltered both in a biofilm and in a sarA mutant. For example, expression of scpA was decreased in a biofilm but elevated in a sarA mutant. Expression of the gene encoding aureolysin (aur) was not altered in a biofilm but was increased in a sarA mutant [19,33]. Expression of the sspABC operon was elevated in a biofilm and a sarA mutant, but because no direct comparison has been made, it is not known whether expression of sspABC is elevated in a sarA mutant even by comparison to a biofilm.

It is clear that overall extracellular protease activity is elevated in sarA mutants [36–38], but to date studies that have attempted to address the issue have concluded that this has relatively little impact on the biofilm-deficient phenotype of a sarA mutant. For instance, mutation of sspA, which encodes the classic “V8” serine protease (SspA), had no impact on biofilm formation in either UAMS-1 or its sarA mutant [33]. Similarly, Valle et al. (2003) concluded that the increased production of proteases in sarA mutants was unlikely to explain their biofilm-deficient phenotype based on the observations that 1) the capacity of a sarA mutant to form a biofilm was not enhanced in the presence of the protease inhibitors α2-macroglobulin or E64, 2) the capacity of the wild-type strains (ISP479C and 15981) to form a biofilm was not reduced by incubation in concentrated supernatants from sarA mutants, and 3) concomitant mutation of sarA and sspA or sarA and the aureolysin gene (aur) failed to enhance biofilm formation.

In contrast to such studies, Boles and Horswill (2008) recently concluded that elevated protease production can be correlated with reduced biofilm formation and that the increased production of extracellular proteases plays an important role in agr-mediated dispersal from an established biofilm. These studies were limited to derivatives of SH1000, which is an rsbU-repaired derivative of 8325-4, and they did not address the potential contribution of extracellular proteases to the biofilm-deficient phenotype of sarA mutants, but...
Figure 5. Impact of *alsSD* expression in a *sarA* mutant. Panel A: Expression of *alsSD* was assessed by qRT-PCR. Results represent the mean ± standard deviation of the results obtained with 3 replicate samples. Panel B: 18-hour culture supernatants were assayed for acetoin production using the Voges-Proskauer assay. Results represent the mean ± standard deviation of 6 replicate experiments. Panel C: Relative levels of biofilm formation were determined using the microtiter plate assay. Results represent the mean ± standard deviation of 16 replicate experiments. Strain designations are the same as in previous figures with the addition of UAMS-1729 (U1729) and UAMS-1730 (U1730), both of which are UAMS-1 *sarA* mutants complemented with pLS50::*alsSD*.

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Figure 6. Production of extracellular nuclease. Results were assessed after overnight incubation of the indicated strains overnight on DNase test agar plates. Numbers refer to UAMS strain designations (Table 1).

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such results nevertheless suggest that previous studies may not have fully defined the impact of protease production in this context. Based on this, we employed a combination of three protease inhibitors in an attempt to collectively limit the activity of all recognized *S. aureus* extracellular proteases. Specifically, E-64, 1,10-phenanthroline, and dichloroisocoumarin (DIC) inhibit the activity of cysteine proteases (e.g. ScpA and SspB), metalloproteases (e.g. aureolysin), and serine proteases (e.g. SspA and the Spl proteases) respectively.

Each inhibitor was added to the medium used in our biofilm assays at a concentration of 1 mM or, in the case of phenanthroline, the highest concentration (10 μM) that did not limit growth (Fig. 8). In fact, addition of these inhibitors both alone or in combination with each other increased growth of the *sarA* mutant to a limited degree at least as defined by the maximum optical density of post-exponential phase cultures (Fig. 8). This suggests that the increased production of proteases may actually limit the growth of an *S. aureus sarA* mutant. Subsequent analysis using skim milk-agar plates confirmed that this combination of protease inhibitors also decreased overall proteolytic activity by comparison to the *sarA* mutant (data not shown). More importantly, at least in the context of this report, inclusion of these inhibitors also enhanced biofilm formation in a *sarA* mutant (*p*<0.001) (Fig. 9A). Although a slight increase in biofilm formation was also observed with the parent strain, analysis of the relative effects confirmed that the impact in a *sarA* mutant significantly exceeded that observed in UAMS-1 (Fig. 9B). These results indicate that the increased production of extracellular proteases also makes an important contribution to the biofilm-deficient phenotype of a UAMS-1 *sarA* mutant.

One possible explanation is that the presence of protease inhibitors increased growth of the *sarA* mutant, particularly with respect to the maximum density observed with post-exponential phase cultures (Fig. 8). However, we observed a similar degree of growth enhancement when each inhibitor was included alone as well as in combination with each other, and this was not the case with respect to biofilm formation. Specifically, E-64 was the only individual inhibitor in which a significant increase in biofilm formation was observed either alone or in combination with DIC or phenanthroline (*p*<0.001) (Fig 10). This suggests that the effect of the inhibitor cocktail on biofilm formation cannot be explained by its impact on growth of the *sarA* mutant in biofilm medium. It also indicates that the cysteine proteases ScpA and/or SspB play an important role. This is in contrast to the results of Valle *et al.* (2003), who concluded that E-64 has no impact on the biofilm-deficient phenotype of a *sarA* mutant. However, the concentration of E-64 employed in these earlier experiments (10 μM) was considerably lower than that used in our experiments (1 mM). At the same time, the impact of E-64 even at the higher concentration was limited by comparison to both the wild-type strain (data not shown) and the *sarA* mutant grown in the presence of all three inhibitors (*p* = 0.012) (Fig. 10), and this clearly suggests that *sarA*-regulated proteases other than ScpA or SspB are also involved.

In an effort to further define the role of specific proteases, we also examined the impact of mutating *sspA* in both UAMS-1 (UAMS-960) and its *sarA* mutant (UAMS-962). In both cases, mutation of *sspA* had no significant impact on biofilm formation (Fig. 9A). Additionally, the enhanced biofilm formation observed with a UAMS-1 *sarA* mutant grown in the presence of all three inhibitors was also observed with a *sarA/spa* double mutant (*p*<0.001) (Fig. 9B). This indicates that the impact of the protease inhibitors on biofilm formation was independent of SspA. This is consistent with the results of Valle *et al.* (2003), who also found that mutation of *spa* had no impact on biofilm formation in a *sarA* mutant.

To further examine this issue, we carried out zymogram analysis using both casein and gelatin gels. This analysis confirmed that both the inhibitor cocktail and E-64 alone or in combination with either DIC or phenanthroline reduced the activity of cysteine proteases to undetectable levels as defined by the sensitivity of our zymogram assays (Fig. 11). In contrast, we could not demonstrate significant inhibition of serine or metalloproteases with DIC or phenanthroline. Moreover, production of SspA was unaffected by the inclusion of DIC in the growth medium (Fig. 11). To the extent that DIC is a specific inhibitor of serine proteases, including SspA, this suggests that the concentration of DIC used in our experiments was below the level required to have a phenotypic effect. We could not determine whether the same was true for the *spl*-encoded serine proteases because we could not detect the activity of these enzymes in either of our zymograms. This is perhaps not surprising since the activity of these proteases was recently shown to be dependent on their binding of specific substrates in a manner analogous to the exfoliative toxins [46,47].

![Figure 7](image.png)

**Figure 7. Effect of nuclease on biofilm formation.** Biofilm formation was assessed using the microtiter plate assay. Results represent the mean ± standard deviation of 24 replicates. Strains designations are the same as in previous figures with the addition of UAMS-1725 and UAMS-1726, which are the *sarA/nuc* mutant complemented with *nuc* and *sarA* respectively. doi:10.1371/journal.pone.0003361.g007
Similarly, we could not demonstrate inhibition of the metalloprotease aureolysin with phenanthroline because we could not detect the activity of this protease even in a \textit{sarA} mutant (Fig. 11). Nevertheless, the fact that the combination of all three proteases inhibitors enhanced biofilm formation to an extent that exceeded that observed with any single or even dual combination of protease inhibitors (Fig. 10) suggests that the concentrations of both DIC and phenanthroline used in these experiments were in fact inhibitory. It is also consistent with the hypothesis that extracellular proteases other than the cysteine proteases ScpA and SspB also make an important contribution to the biofilm-deficient phenotype of a \textit{sarA} mutant. To the extent that this effect was independent of \textit{sspA} (see above), this suggests that the \textit{spl}-encoded serine proteases and/or aureolysin may be particularly important in this regard. Although confirmation of this hypothesis will require examination of UAMS-1 \textit{sarA/aur} and \textit{sarA/spl} mutants, this is consistent with the results of Boles and Horswill (2008), who detected increased levels of serine proteases in the effluent from \textit{S. aureus} biofilms and demonstrated that concomitant mutation of \textit{aur} and \textit{spl} resulted in an enhanced capacity to form a biofilm at least in the 8325-4 strain SH1000.

While the combination of protease inhibitors enhanced biofilm formation in a \textit{sarA} mutant, it did not restore it to wild-type levels (Fig. 9A). One explanation for this is that the concentration of inhibitors used in our experiments was sufficient to limit protease activity in a \textit{sarA} mutant but not sufficient to restore it to wild-type levels. As discussed above, this is particularly true with respect to DIC and phenanthroline. It was not possible to use either of these inhibitors at a higher concentration either due to issues related to limited solubility in biofilm medium (DIC) or inhibition of growth (phenanthroline), so addressing this possibility will also require detailed analysis of relevant protease mutants. Nevertheless, the results discussed above suggest that the increased production of both extracellular nuclease and multiple extracellular proteases contribute to the biofilm-deficient phenotype of an \textit{S. aureus} \textit{sarA} mutant. However, they also demonstrate that the biofilm-defect in a \textit{sarA} mutant cannot be explained by its impact on either nuclease or proteases alone. To determine whether these effects might be cumulative, we also examined whether inclusion of protease inhibitors had an impact on biofilm formation with a \textit{sarA/nuc} mutant. The results confirmed that the effect was cumulative in that biofilm formation was enhanced in a \textit{sarA/nuc} mutant in the presence of protease inhibitors by comparison to both a \textit{sarA} mutant in the presence of inhibitors (\textit{p}<0.001) and a \textit{sarA/nuc} mutant in the absence of inhibitors (\textit{p}<0.001) (Fig. 9A). In fact, in the presence of the inhibitor cocktail, the \textit{sarA/nuc} mutant exhibited a level of biofilm formation approaching that of the UAMS-1 parent strain.

**Conclusions**

Taken together, our results provide important clues with respect to explaining the biofilm-deficient phenotype of a \textit{S. aureus} \textit{sarA} mutant, and there may in fact be a common theme that ties them all together. For instance, a recent report demonstrated that mutation of \textit{cidA}, which encodes a regulator of murein hydrolase activity, results in reduced release of extracellular DNA and a reduced capacity to form a biofilm [35]. The \textit{cidA} gene is part of the \textit{cidR} regulon, which also includes \textit{alsSD} [44]. Our results confirmed that mutation of \textit{alsSD} in UAMS-1 results in a stationary-phase survival defect manifested as cell death in the absence of cell lysis. This suggests that mutation of \textit{alsSD} may limit the release of extracellular DNA as well as limit the capacity to...

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**Figure 8. Effect of protease inhibitors on growth.** Growth of the UAMS-929 \textit{sarA} mutant in biofilm medium without protease inhibitors (\textbullet) was compared to growth in the same medium containing 1 mM DIC (\texttriangle), 1 mM E-64 (\textcircle), 10 μM 1,10-phenanthroline (\textasteriskcentered) or a cocktail containing all three inhibitors (\texttimes). Growth was monitored at hourly intervals for 10 hours. Results indicate the OD\textsubscript{600±} standard deviation of 3 replicates.

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produce acetoin and maintain pH homeostasis. In fact, one explanation for our results is that mutation of alsSD limits biofilm formation because it limits the availability of extracellular DNA but that this effect is modest by comparison to the impact of sarA on the production of extracellular nuclease. Such a scenario might also provide an alternative explanation for why restoration of alsSD expression in a sarA mutant failed to restore biofilm formation since what little extracellular DNA would be available would presumably be degraded due to the increased production of nuclease.

How the production of proteases might fit into this scenario remains undefined. The production of active extracellular nuclease does require protease processing [48], but if this were a defining characteristic of the biofilm-deficient phenotype of a sarA mutant, then mutation of nuc and inclusion of protease inhibitors would not be expected to have a cumulative effect since mutation of nuc eliminates the production of nuclease in any form. This suggests that nuclease and protease have independent effects that remain to be defined but are perhaps related to the attachment vs. accumulation phases of biofilm formation. Given our experimental approach using a cocktail of protease inhibitors, it also remains to be determined exactly what proteases are involved, how they interact with each other, what S. aureus proteins are the most relevant targets, and what role these targets play in biofilm formation. In this respect it should be noted that biofilm formation in UAMS-1 and other clinical isolates is enhanced by coating the substrate with plasma proteins [12] and that, at least in UAMS-1, it is not dependent on PIA production [19]. Moreover, we have demonstrated that the increased production of proteases in sarA mutants results in a decreased capacity to bind host proteins including fibronectin [36]. Taken together, these results suggest that the negative impact of proteases on biofilm formation may be multifactorial and involve both attachment and accumulation. A recent report also demonstrated that induction of agr expression leading to increased production of multiple proteases may also serve as a specific means of dispersal from an established biofilm [40].

While our experimental focus is on the role of sarA in biofilm formation, our results may also have broader implications for S. aureus regulatory circuits. For instance, sarA represses production of

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**Figure 9.** Effect of protease inhibitors on biofilm formation. Panel A: The microtiter plate assay of *in vitro* biofilm formation was performed with bacteria grown in biofilm medium (gray) or biofilm medium containing the inhibitor cocktail (black). Results represent the mean ± standard deviation of 24 replicates. Panel B: Results of the biofilm assay shown in panel A represented as the ratio of biofilm formation in the presence of the inhibitor cocktail/biofilm formation in the absence of inhibitors.

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that role in this regard. Together with our results, this would suggest concluded that
many S. aureus exoproteins including nuclease and multiple proteases [33,36] while the accessory gene regulator (agr) has the opposite effect [25,40]. While there are reports concluding that reduced expression of agr can be correlated with a reduced capacity to form a biofilm [18,49–51], most studies have concluded that agr expression limits rather than enhances biofilm formation [5,12,52]. In fact, it has been proposed that induction of agr expression may serve as a specific means of dispersal from an established biofilm [5,12,22,40].

One report suggested that the negative impact of agr on biofilm formation may be due to the increased production of delta-toxin, which is a phenol-soluble modulin (PSM) proposed to act as a surfactant to limit cellular accumulation [5,53]. The alternative explanation is that expression of agr results in the increased production of both extracellular nuclease and protease(s) [25]. This would presumably promote detachment and release from a mature biofilm based on degradation of the relevant S. aureus adhesins and/or extracellular matrix components including extracellular DNA. Indeed, Boles and Horwill (2008) recently concluded that agr-mediated protease production plays a primary role in this regard. Together with our results, this would suggest that sarA and agr have independent and opposite effects on biofilm formation, with the ultimate impact being dependent on the relative contribution of each with respect to the other.

In this scenario, S. aureus strains that express agr at high levels would presumably have a reduced capacity to form a biofilm. This is consistent with our observation that the 8325-4 strain RN6390, which by comparison to most clinical isolates including UAMS-1 expresses agr at high levels and produces elevated amounts of nuclease and extracellular proteases [36], has a limited capacity to form a biofilm [12]. It is also consistent with the observation that mutation of agr enhances biofilm formation in RN6390 [12]. This effect is reversed by concomitant mutation of sarA [12], which suggests that the impact of sarA is epistatic to agr at least in this context.

It is unclear whether this has any relevance with isolates other than RN6390, which as noted above has specific characteristics that distinguish it from clinical isolates of S. aureus. However, a recent report concluded that most community-acquired MRSA (CA-MRSA) isolates express agr at higher levels than their healthcare-associated MRSA (HA-MRSA) counterparts [53]. Our studies done with a USA300 CA-MRSA isolate suggest that this does not preclude biofilm formation at least in this strain [52], but there are reports demonstrating a general inverse relationship between the level of agr expression and biofilm formation [5]. Together with the fact that agr expression is also correlated with the increased production of S. aureus extracellular proteins and PSMs, the latter having both surfactant-like and anti-phagocytic properties [53], this could perhaps explain why CA-MRSA isolates often cause acute infections while HA-MRSA tend to cause chronic infections that are more likely to have a biofilm-associated component [54].

Finally, our previous studies characterizing the sarA and biofilm regulons in UAMS-1 [19,33] identified 43 genes that were differentially expressed both in a biofilm and in a sarA mutant [12,33]. Of the four possible scenarios, 17 genes were expressed at lower levels in both a biofilm and a sarA mutant, 6 genes were expressed at higher levels in both a biofilm and a sarA mutant, 17 genes were expressed at lower levels in a biofilm and higher levels in a sarA mutant, 3 genes were expressed at lower levels in a biofilm and lower levels in a sarA mutant. Given the complex and interactive nature of regulatory circuits in S. aureus, it is difficult to predict which of these scenarios would be most important, and we certainly do not preclude the need to investigate additional genes in the sarA/biofilm regulon. However, our results suggest that the inability of a sarA mutant to repress production of specific extracellular proteins, including nuclease and multiple proteases, play a particularly important role in that regard. A detailed understanding of these processes is important given the role of biofilms not only in the development of many forms of S. aureus infection but also with respect to their impact on the ability to effectively treat these infections.

Materials And Methods

Bacterial strains and growth conditions

The strains utilized in this study are listed in Table 1. All strains were maintained as stock cultures at −80°C in tryptic soy broth (TSB) containing 25% (v/v) glycerol. For each experiment, the relevant strains were retrieved from cold storage by plating on

Figure 10. Biofilm formation in the presence of individual inhibitors alone and in combination with each other. Biofilm formation in the UAMS-929 sarA mutant was assessed in the absence of protease inhibitors (Φ), the presence of the protease inhibitor cocktail (CT), or in presence of individual inhibitors alone and in paired combinations with each other. Inhibitor designations are E-64 (E), DIC (D) and phenanthroline (P). Results represent the mean ± standard deviation of 24 replicates.
doi:10.1371/journal.pone.0003361.g010
tryptic soy agar (TSA) with appropriate antibiotic selection. Antibiotics were used at the following concentrations: kanamycin (Kan; 50 \( \mu \)g per ml), neomycin (Neo; 50 \( \mu \)g per ml), chloramphenicol (Cm; 10 \( \mu \)g per ml), and erythromycin (Erm 5 \( \mu \)g per ml). To ensure that the results of phenotypic assays were consistent, all assays other than nuclease production (see below) were done using TSB supplemented with 0.5% glucose and 3.0% sodium chloride (biofilm medium) without antibiotic selection as previously described [12].

Mutagenesis

Mutagenesis of alsSD and nuc was done as previously described [52] using the pKOR1 mutagenesis system [45]. According to the annotation for the MRSA252 genome, which was previously shown to be the most closely related of the sequenced strains to UAMS-1 [21], the specific open-reading frames (ORFs) targeted in these experiments were SAR2297 (alsS), SAR2296 (alsD), and SAR0947 (nuc). The corresponding ORFs in the N315 genome are SA2008/SA2007 (alsSD) and SA0746 (nuc). The oligonucleotide primers used for mutagenesis are listed in Table 2. In UAMS-1489, the deleted region started 650 bp downstream of the alsS start codon and ended 182 bp downstream of the alsD stop codon (primers alsSDMut1FattB1, alsSDMut1RSacII, alsSD2FSacII, alsSD2RattB2). The corresponding ORFs in the N315 genome are SA2008/SA2007 (alsSD) and SA0746 (nuc). The oligonucleotide primers used for mutagenesis are listed in Table 2. In UAMS-1489, the deleted region started 650 bp downstream of the alsS start codon and ended 182 bp downstream of the alsD stop codon (primers alsSDMut1FattB1, alsSDMut1RSacII, alsSD2FSacII, alsSD2RattB2).

Complementation of alsSD, nuc and sarA mutations

Complementation of the alsSD mutation was done by cloning the region spanning 431 bp upstream and 118 bp downstream of the alsSD operon (primers alsSDproF and alsSD downstream KpnI) into the E. coli/S. aureus shuttle vector pLI50 [53]. Complementation of the nuc mutation was done by cloning the region spanning 433 bp upstream of 277 bp downstream of the nuc gene (primers nuc comp-
F and nuc comp-R) into pLI50. Complementation of the sarA mutation was done as previously described [36]. In all cases, complementing plasmids were first introduced into the corresponding UAMS-1 mutants using Φ11 as previously described [36].

Phenotypic assays

The Voges-Proskauer assay was used to assess the production of acetoin and 2,3-butanediol as previously described [44]. Briefly, cultures were grown for 18 hrs in biofilm medium (TSB supplemented with 0.5% glucose and 3.0% NaCl) with constant shaking before harvesting cell-free supernatants by centrifugation. The assay was done in 96-well microtiter plates by adding 50 μl of 0.3% creatine and 60 μl of 5% alpha-naphthol freshly prepared in 100% ethanol to 120 μl of fresh culture supernatant. After gentle mixing, 30 μl of 40% KOH was added to the reaction mixture, which was then incubated at room temperature for 30–45 min with occasional mixing. Results were assessed by measuring absorbance at 540 nm.

Biofilm formation was assessed in vitro using the static, microtiter plate biofilm assay as previously described [12]. To assess stationary-phase survival, overnight cultures of each S. aureus strain were grown in TSB with appropriate antibiotic selection and then used to inoculate 21 ml of NZY broth (Fisher Scientific, St. Louis, MO) containing 35 mM glucose to an OD600 of 0.05. Flasks were loosely capped and grown at 37°C with constant shaking. After 8, 24, 48, and 72 hrs, the optical density (OD600) of each culture was determined and an aliquot was removed to determine viable count by plating on TSA.

Nuclease activity was then assessed by overlaying the agar with 1N HCl to precipitate undigested DNA and define the zone of clearance around each strain. Overall protease activity was assessed using skim milk agar as previously described [40], the only difference being that we analyzed standardized culture supernatants after 15-fold concentration using Centricon YM-3 filter units (Millipore, Bedford, MA). The activity of specific proteases was assessed by zymography using Ready Gel Zymogram Gels containing gelatin or casein (BioRad Laboratories, Hercules, CA). For casein gels, supernatants were analyzed without further processing. For gelatin gels, supernatants were first concentrated as discussed above. Samples in both cases were loaded

Table 1. Strains used in this study.

<table>
<thead>
<tr>
<th>Strain</th>
<th>Description</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>UAMS-1</td>
<td>MSSA, osteomyelitis isolate</td>
<td>Gillaspy et al., 1995</td>
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<tr>
<td>UAMS-929</td>
<td>UAMS-1sarA-kan</td>
<td>Blevins et al., 2002</td>
</tr>
<tr>
<td>UAMS-960</td>
<td>UAMS-1spc-tet</td>
<td>Blevins et al., 2002</td>
</tr>
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<td>UAMS-962</td>
<td>UAMS-929spc-tet</td>
<td>Blevins et al., 2002</td>
</tr>
<tr>
<td>UAMS-969</td>
<td>UAMS-929(pLI50:sarA)</td>
<td>Blevins et al., 2002</td>
</tr>
<tr>
<td>UAMS-1300</td>
<td>UAMS-929ΔalsSD</td>
<td>This study</td>
</tr>
<tr>
<td>UAMS-1471</td>
<td>UAMS-1Δnuc</td>
<td>This study</td>
</tr>
<tr>
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<td>UAMS-929Δnuc</td>
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<td>UAMS-1ΔalsSD</td>
<td>This study</td>
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<td>UAMS-1471 (pLI50:nuc)</td>
<td>This study</td>
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<td>UAMS-929 (pLI50:alsSD)</td>
<td>This study</td>
</tr>
<tr>
<td>UAMS-1730</td>
<td>UAMS-929 (pLI50:alsSD)</td>
<td>This study</td>
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</table>

doi:10.1371/journal.pone.0003361.t001

Table 2. Primers used in this study.

<table>
<thead>
<tr>
<th>Primer or Probe</th>
<th>Oligonucleotide Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>alsSD Mut1FattB1</td>
<td>GGG GAC AAG TTT GTA CAA AAA AGC AGG CTC ACA ACA ATC AAT CCA ACA TCC C</td>
</tr>
<tr>
<td>alsSD Mut1RSaclI</td>
<td>ATC GTA GCA GCG GTC AGC ACT AGA ACT TCT CAT ACC</td>
</tr>
<tr>
<td>alsSD 2FscII</td>
<td>ATC GAT CCG CGG ATA TGC AAG TAC TAA ATT CG</td>
</tr>
<tr>
<td>alsSD 2RattB2</td>
<td>GGG GAC CAC TTT GTA CAA AAA AGC TGG GTA TAA ATA AAT CCC CTC ACT ACC G</td>
</tr>
<tr>
<td>nuc Mut1FattB1</td>
<td>GGG GAC AAG TTT GTA CAA AAA AGC AGG CTC GTA CAA TGA CTC TAA GTC AGT CTC ACC</td>
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<tr>
<td>nuc Mut1RsacII</td>
<td>GGA CTC CCG CGG CCA AAT ACT GAT AGC CAT CCC T</td>
</tr>
<tr>
<td>nuc Mut2FscII</td>
<td>GGA CTC CCG CGG CCA AAT ACT GAT AGC CAT CCC T</td>
</tr>
<tr>
<td>nuc Mut2RattB2</td>
<td>GGG GAC CAC TTT GTA CAA AAA AGC TGG GTA CTC CTC TAA TGA TTT GTA TCC</td>
</tr>
<tr>
<td>alsSD proF</td>
<td>CAT TCA TTT ATA TCT ATT TCC CTC T</td>
</tr>
<tr>
<td>alsSD downstream Kpnl</td>
<td>GGA CCT GGT ACC CTA TGA CAA CCA TGC TTA ACC G</td>
</tr>
<tr>
<td>nuc comp-F</td>
<td>ACT TGG CTA AAG CTA CTT GCA AGG</td>
</tr>
<tr>
<td>nuc comp-R</td>
<td>TAA CTC ACA TTT TTC TTC ACG CTC</td>
</tr>
<tr>
<td>alsSD probe</td>
<td>CAT CTC TTT CAT AGC CCT CT TTA TGG CCG</td>
</tr>
<tr>
<td>alsSD RTA</td>
<td>AAG GTT TAC GAG TTA CTA ATC AGG</td>
</tr>
<tr>
<td>alsSD RTS</td>
<td>AAT TTA CAG GTA TAT CAA TTA ATA CTG G</td>
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<tr>
<td>gyrB2 probe</td>
<td>CCG CCA CGG CCG AAT TTA CCA CCA</td>
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<tr>
<td>gyrB RTA</td>
<td>CCA ACA CCA TGT AAA CCA CCA GAT</td>
</tr>
<tr>
<td>gyrB RTS</td>
<td>AGT AAC GGA TAA CAG AGC TGG TA</td>
</tr>
</tbody>
</table>

doi:10.1371/journal.pone.0003361.t002

To assess relative levels of alsSD expression, total bacterial RNA was extracted using the Qiagen RNeasy Mini Kit as previously described [52]. Quantitative, real-time RT-PCR (qRT-PCR) was performed using TaqMan probes corresponding to the alsSD gene (Table 2).

Protease inhibitors

In experiments employing E-64 (Fisher Scientific, St. Louis, MO), and dichloroisoucynamide (DIC) protease inhibitors (Sigma Chemical Co., St. Louis, MO), each inhibitor was dissolved in biofilm medium at a 1 mM concentration. For experiments employing 1-10-phenanthroline (Fisher Scientific, St. Louis, MO), the concentration was reduced to 10 μM because higher concentrations inhibited growth. Subsequent experiments confirmed that these concentrations did not inhibit growth either alone or in combination with each other.

RNA isolation and qRT-PCR analysis

To assess relative levels of alsSD expression, total bacterial RNA was isolated using the Qiagen RNeasy Mini Kit as previously described [52]. Quantitative, real-time RT-PCR (qRT-PCR) was then performed [33] using alsSD-specific primers and a correponding TaqMan probe (Table 2). Results were standardized by comparison to the results obtained with the same samples using primers and a TaqMan probe corresponding to the gyrB gene (Table 2).

Statistical analysis

Statistical comparisons were done using the Student’s t-test or, where appropriate, the Mann Whitney Rank Sum Test as formatted in SigmaStat Statistical Software Version 2 (SPSS Inc., Chicago, IL). Because multiple comparisons were made within each data set, statistical significance, along with the corresponding p value, is noted in the text rather than within each figure.

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Author Contributions

Conceived and designed the experiments: LHT JEC LNS KEB MS. Performed the experiments: LHT JEC LNS KEB. Analyzed the data: LHT JEC LNS KEB MS. Contributed reagents/materials/analysis tools: MS. Wrote the paper: LHT JEC KEB MS.

References


